

Microbial Diversity Using Gene-targeted Pyrosequencing

Design style of high-throughput data



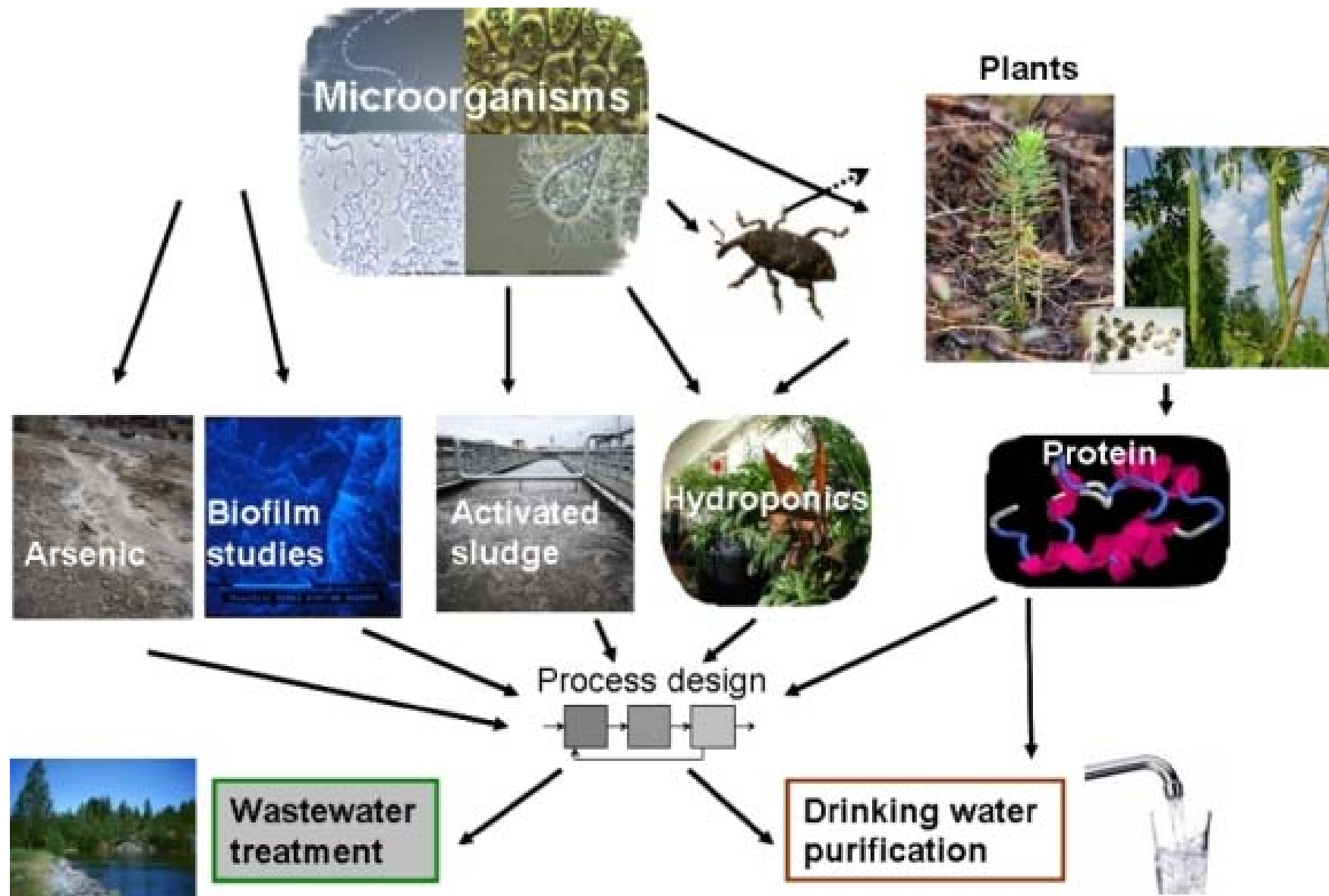
Tae Kwon Lee

WCU Center for Green Metagenomics



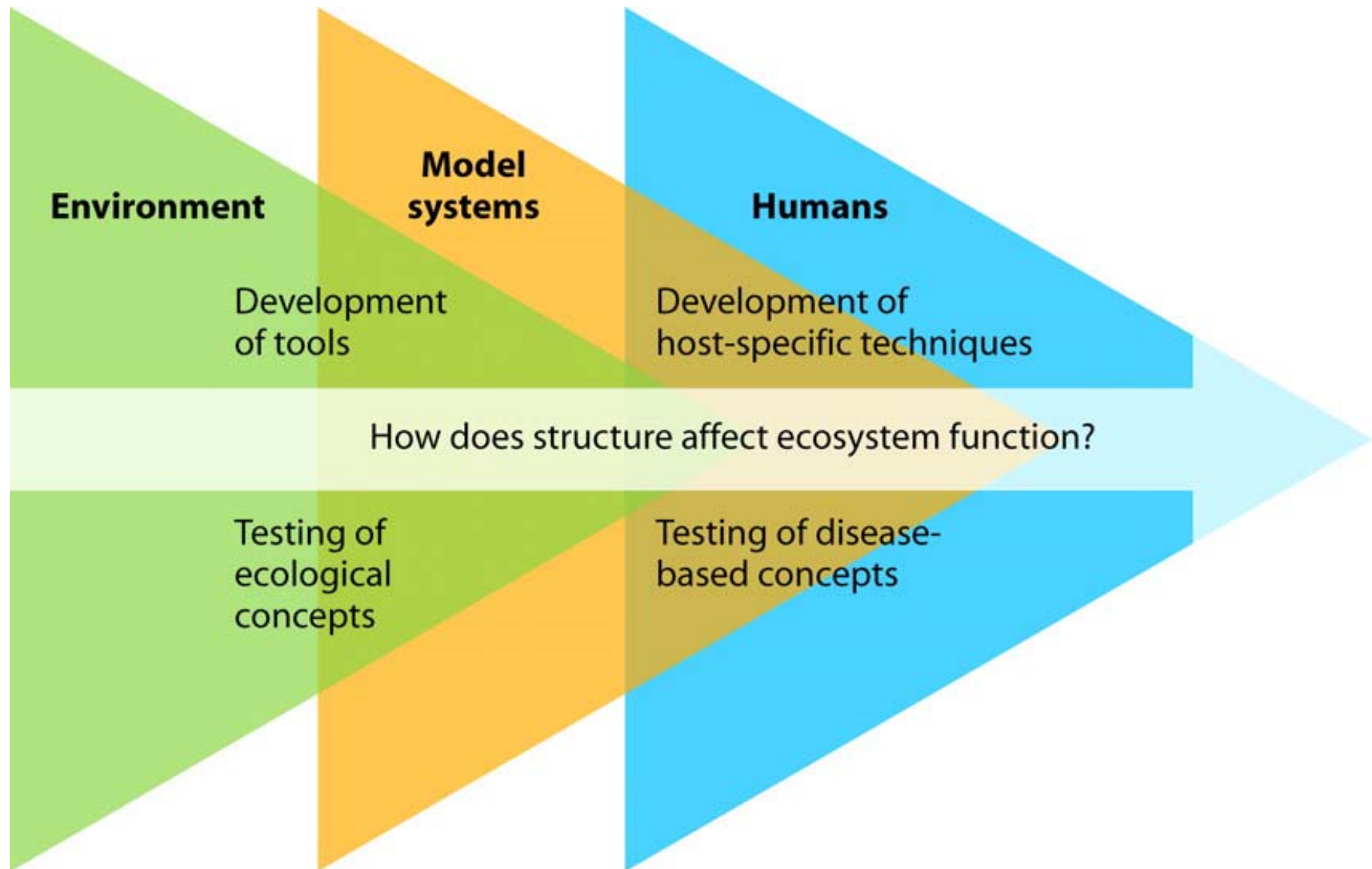
**To Bore
No More**

- 1. Can we determine the true microbial diversity (phylogenetically and genetically)?**
- 2. Is there a core microbiome?**
- 3. Does a structure impact ecological function? If so, how?**

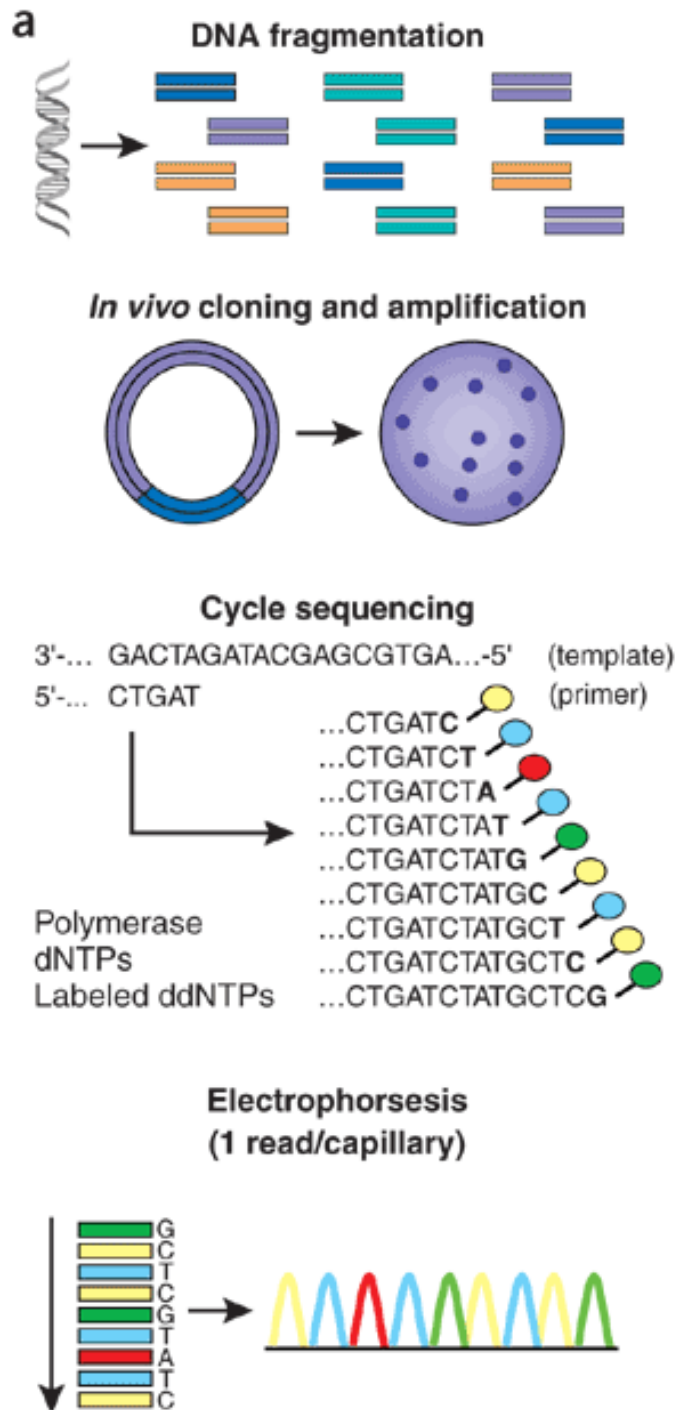


2011, Environmental microbiology, KTH biotechnology

Human Microbiome Project

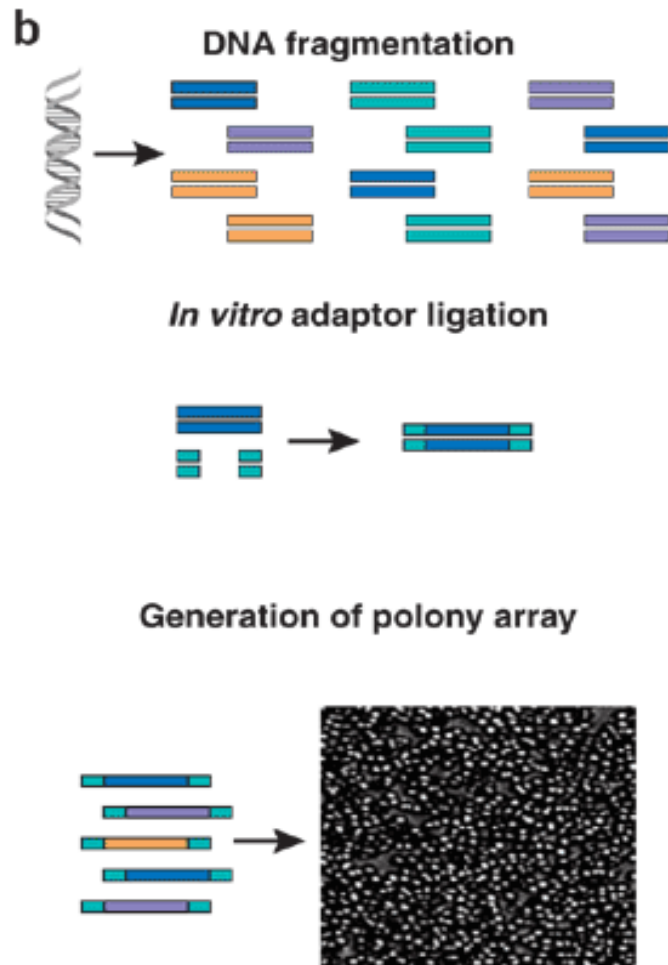


2010, Microbiol. Mol. Biol. Rev. Robinson et al.

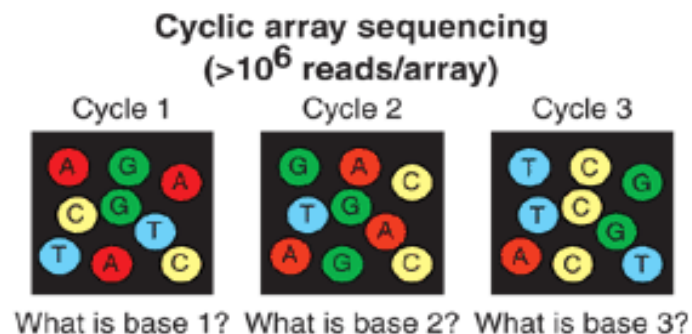


Shotgun Sanger Sequencing

2008, Nature Biotechnology, Shendure & Ji



Next Generation Sequencing



2008, Nature Biotechnology, Shendure & Ji

2nd NGS



The New
GS Junior



MiSeq

3rd NGS



ion torrent
△ ★ ▲ ○ × □ + ≈

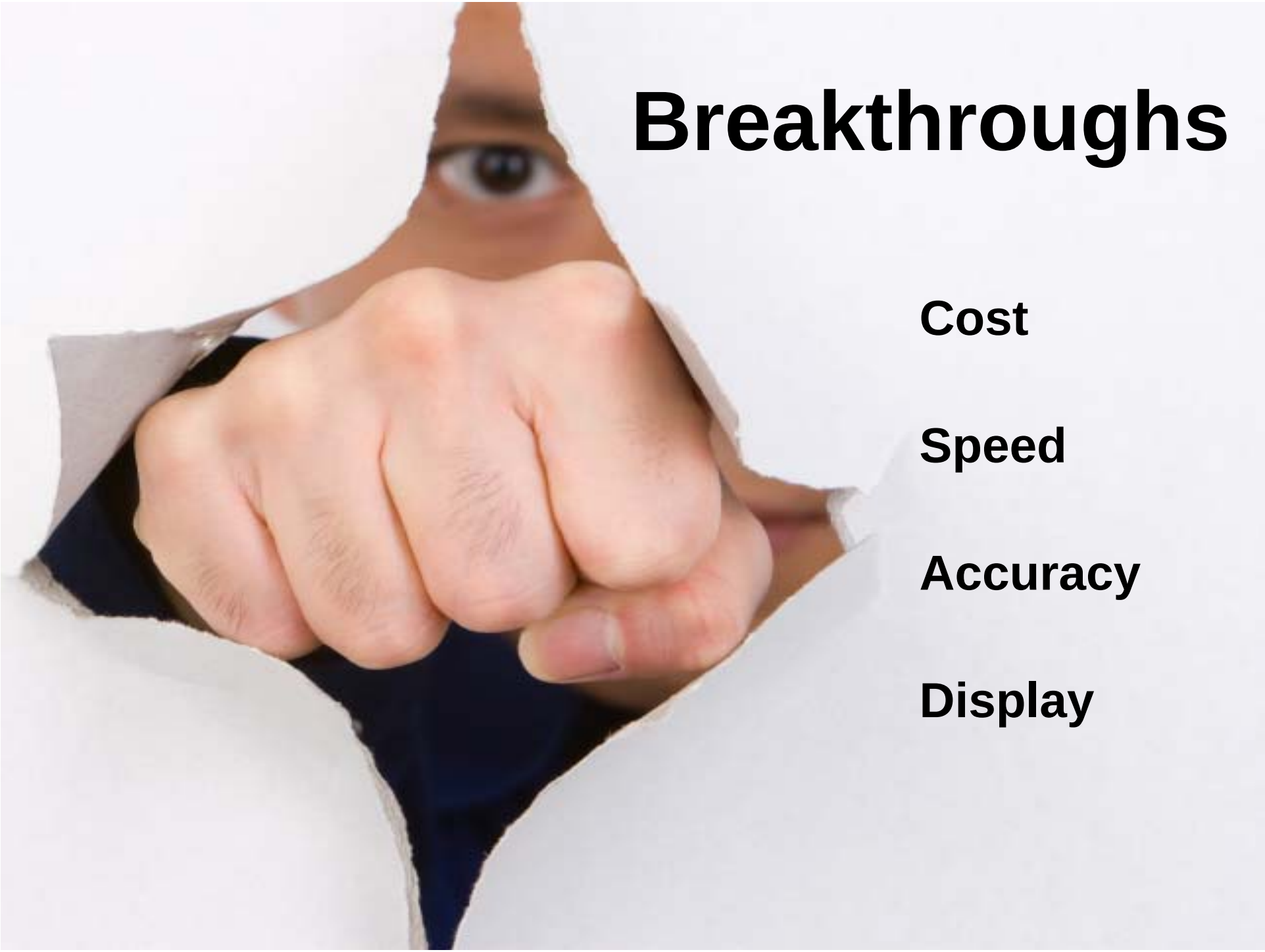


PACBIO RS

Drowned in next generation sequencing data

HELP!



A hand is shown tearing through a piece of white paper. The hand is positioned in the center-left of the frame, with fingers curled as if pulling the paper apart. Behind the paper, a person's face is visible, with one eye looking directly at the camera. The background is a solid light gray.

Breakthroughs

Cost

Speed

Accuracy

Display

- 1. Can we determine the true microbial diversity (phylogenetically and genetically)?**
- 2. Is there a core microbiome?**
- 3. Does structure impact ecological function? If so, how?**



3

Steps of

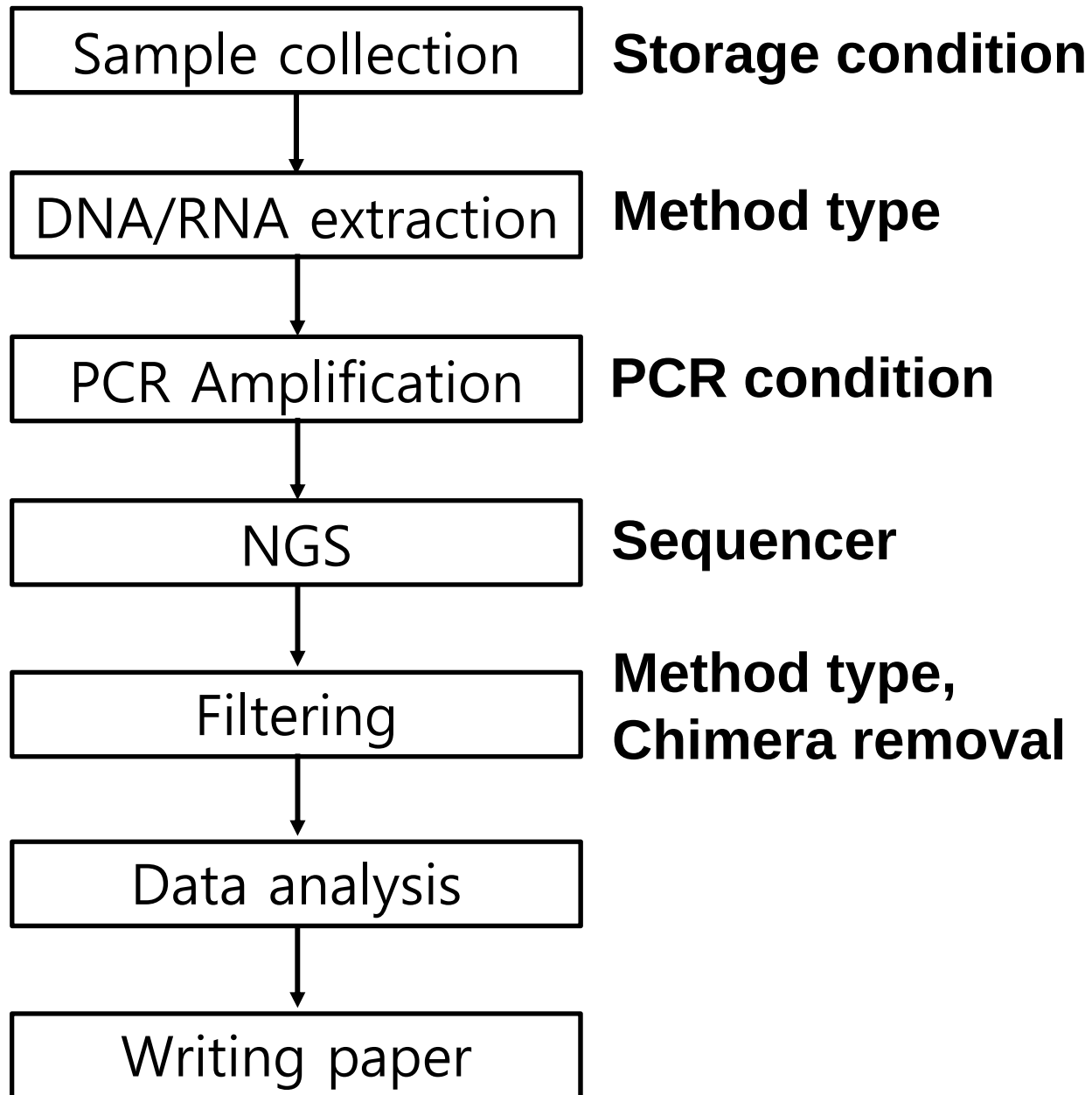
Microbial analysis

1. **Sample preparation & filtering**
2. **Microbial structure & diversity**
3. **Interaction with function**

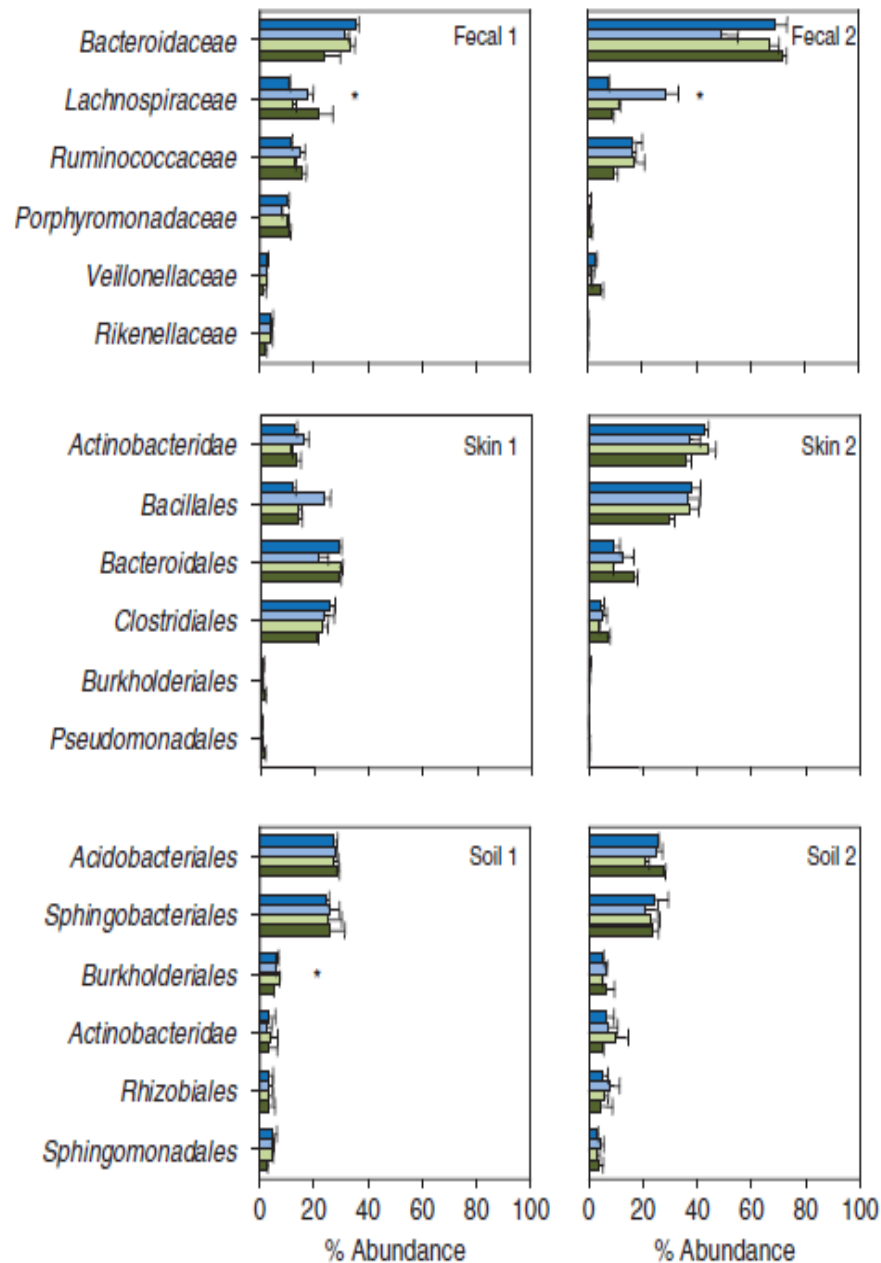


Step 1

Sample preparation and filtering



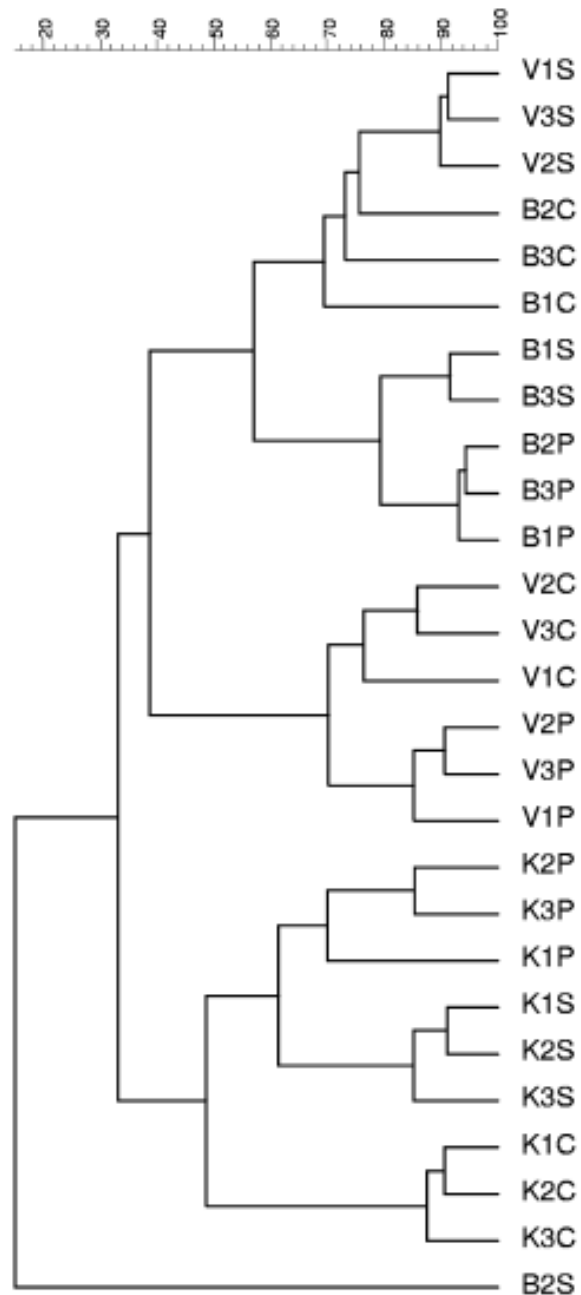
**Sample
preparation
& filtering**



**Less effect of
storage condition**

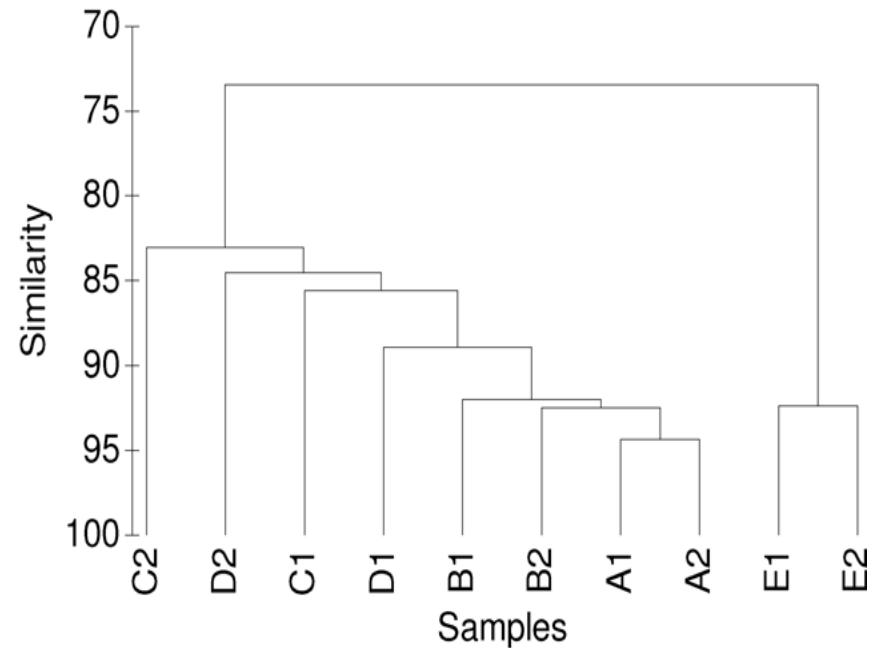
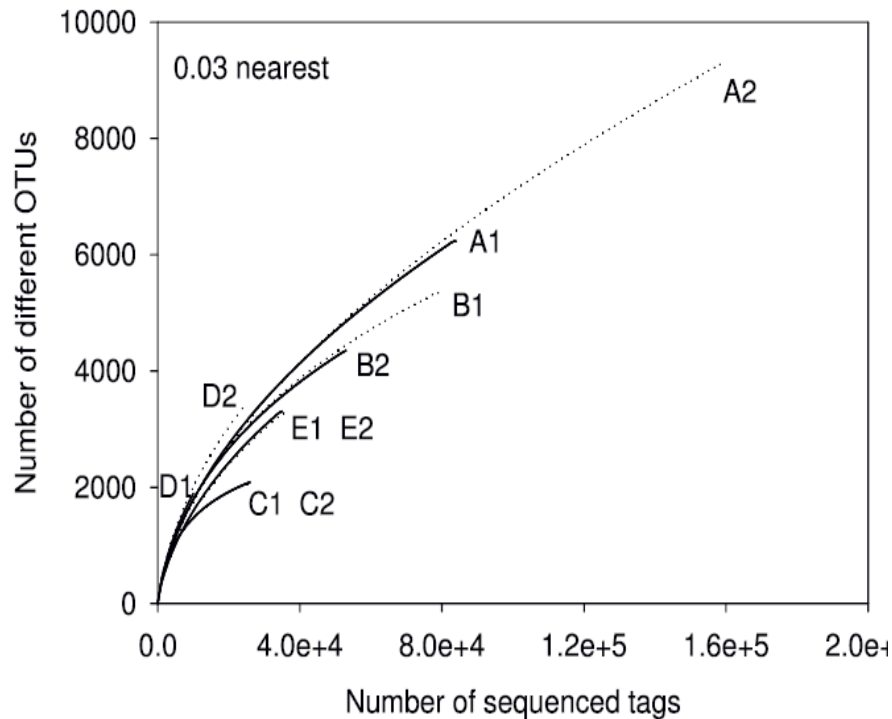
For RNA, use RNALater

2010, FEMS Microbiol Lett. Lauber et al.



**Use of same
extraction method**

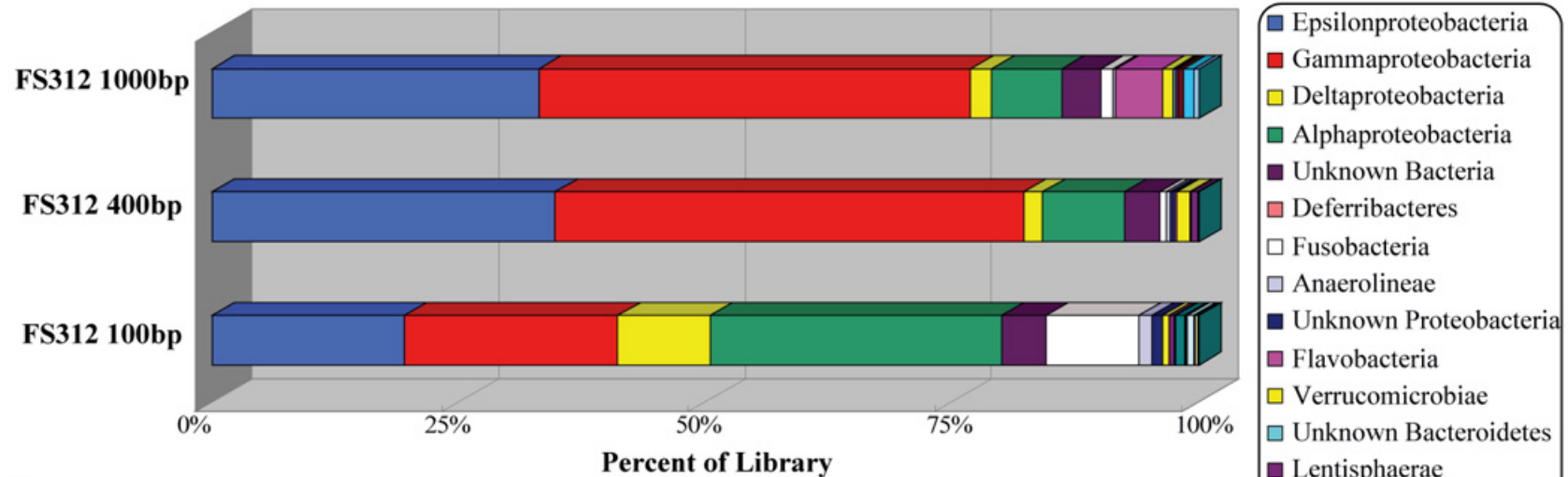
2010, Appl. Environ. Microbiol. Inceoglu et al.



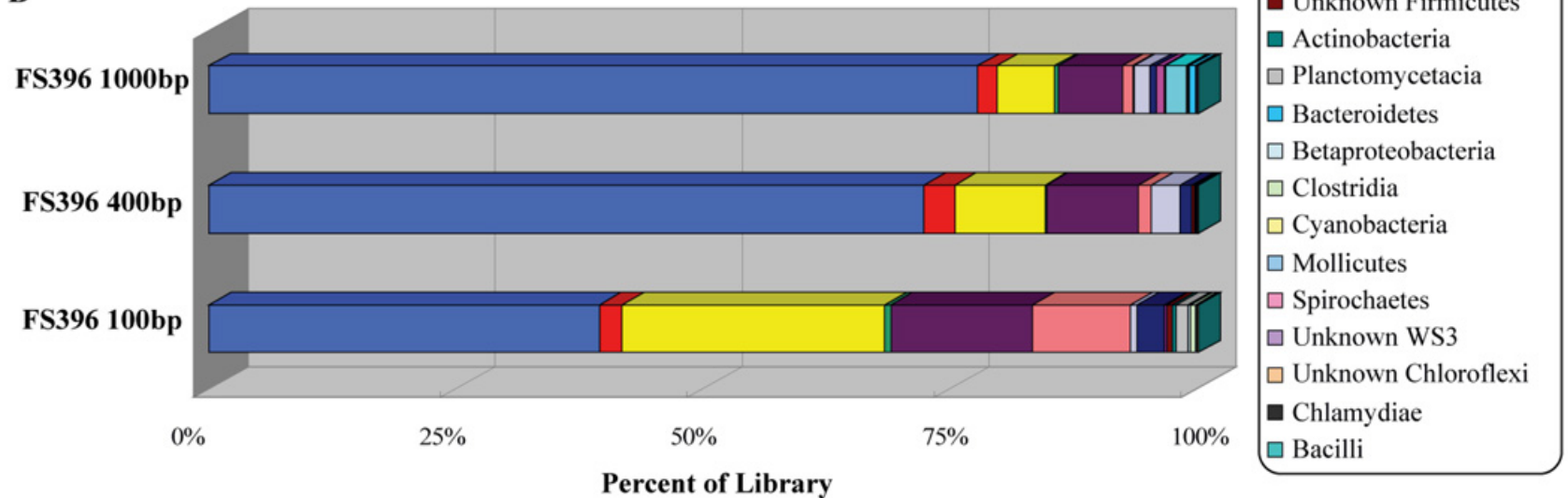
Less effect of Dilution/Cycle

Much effect of Polymerase

A

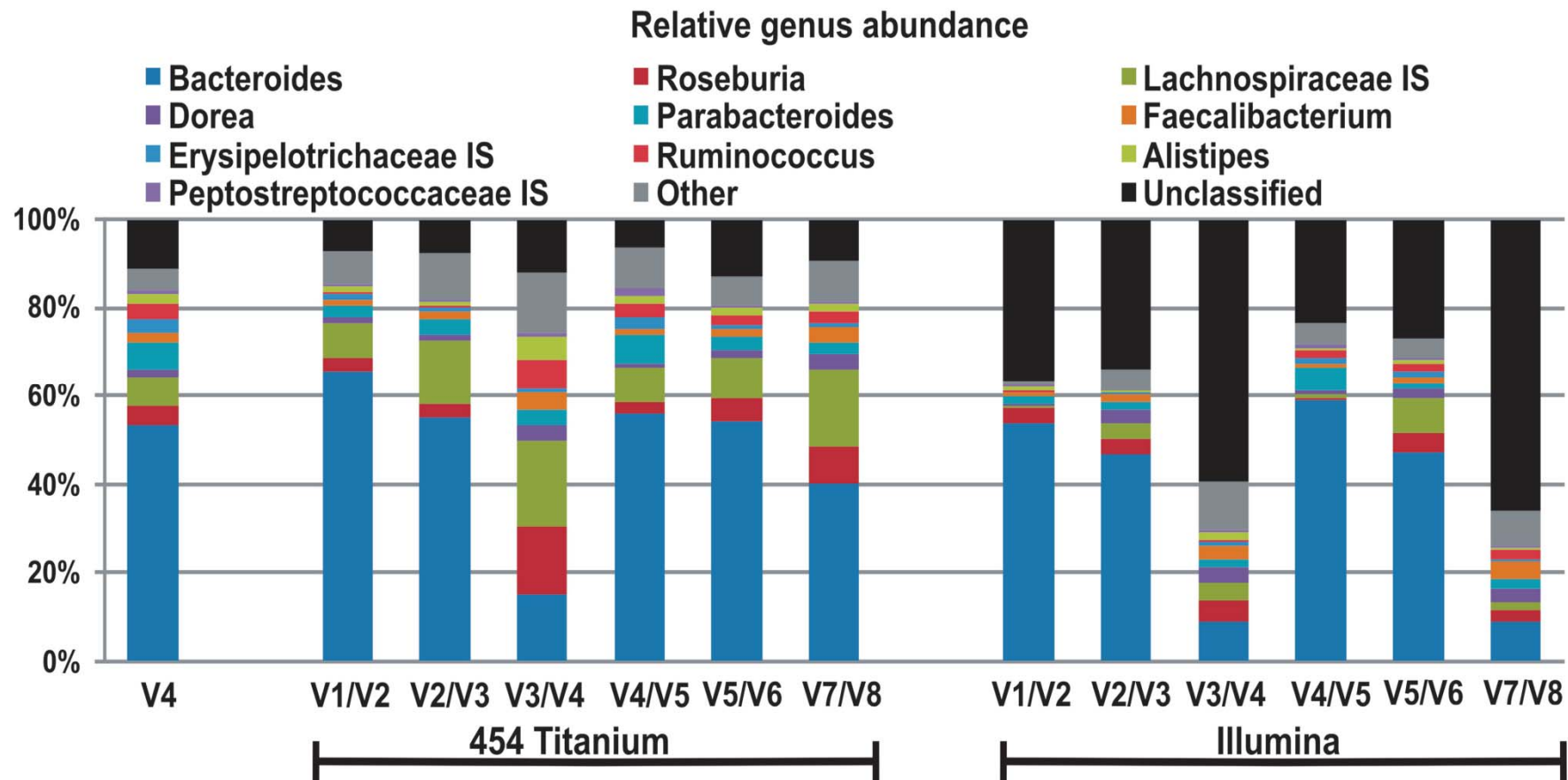
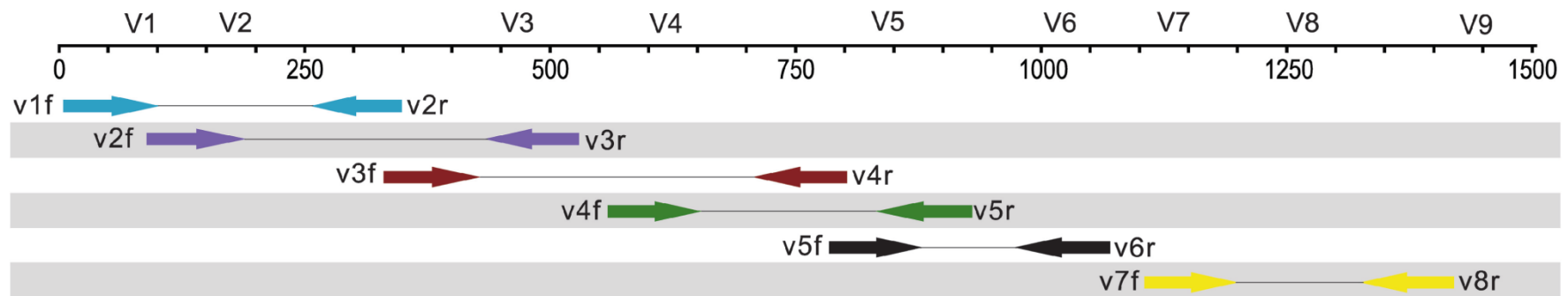


B



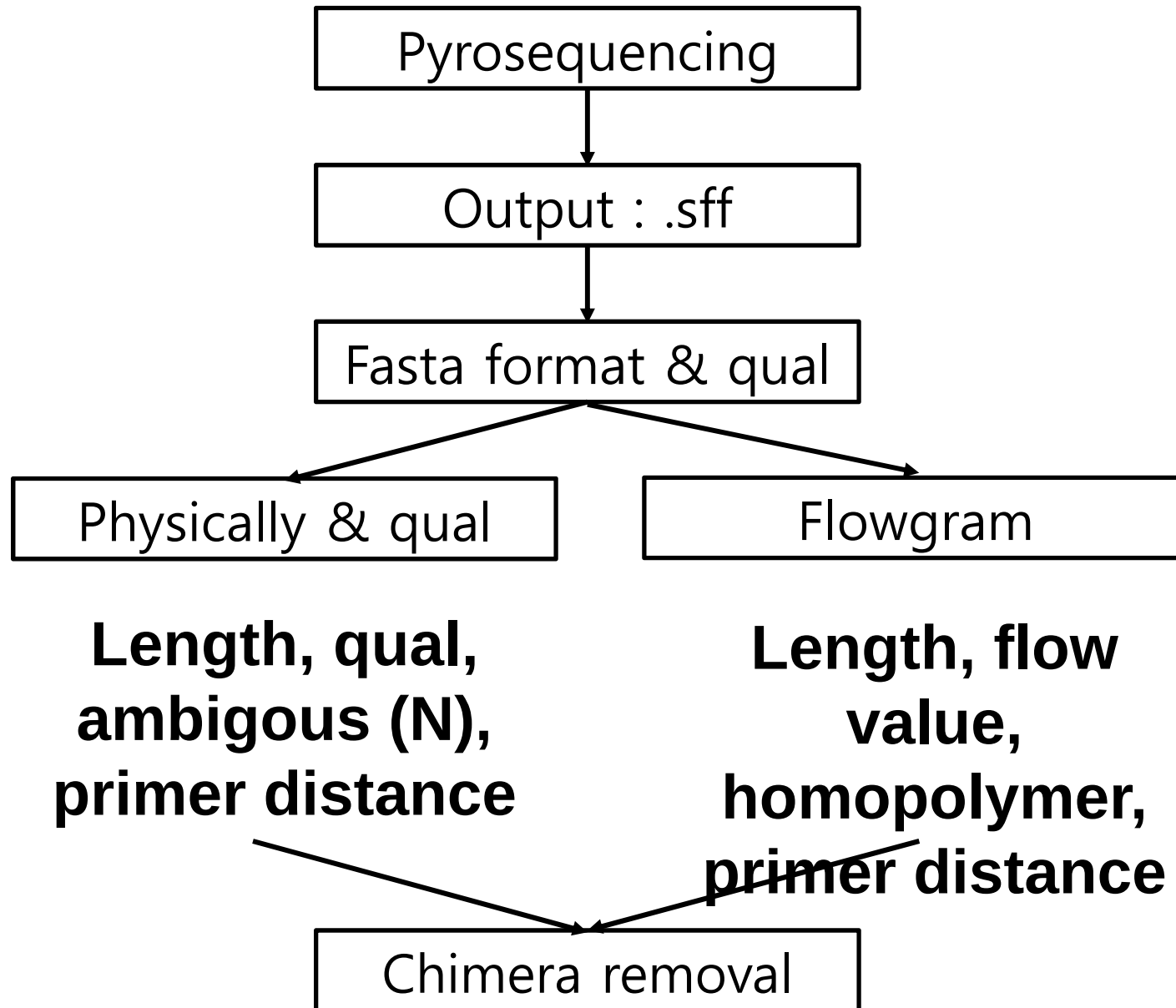
Same primer set and length

2009, Environ. Microbiol. Huber et al.

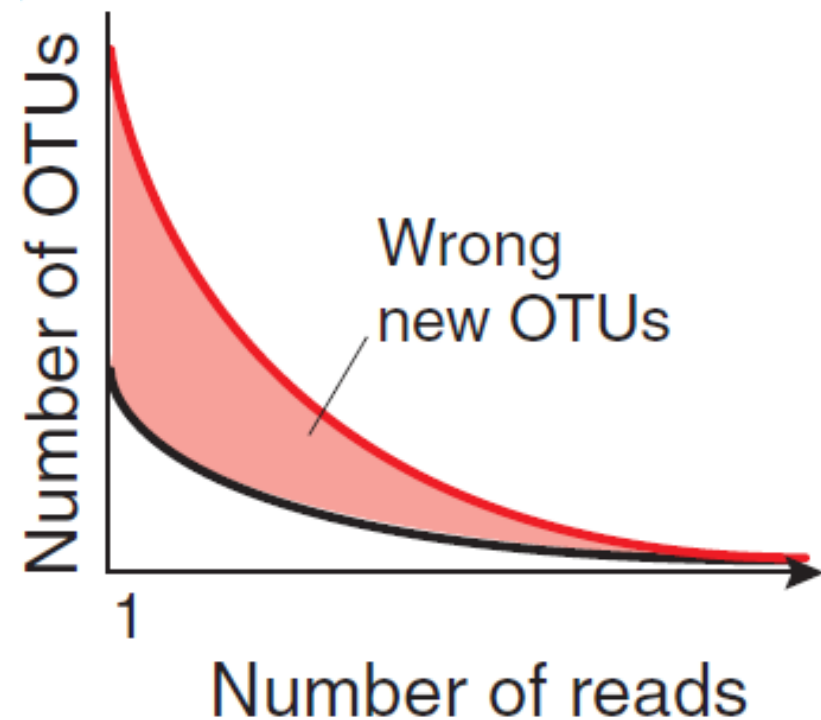
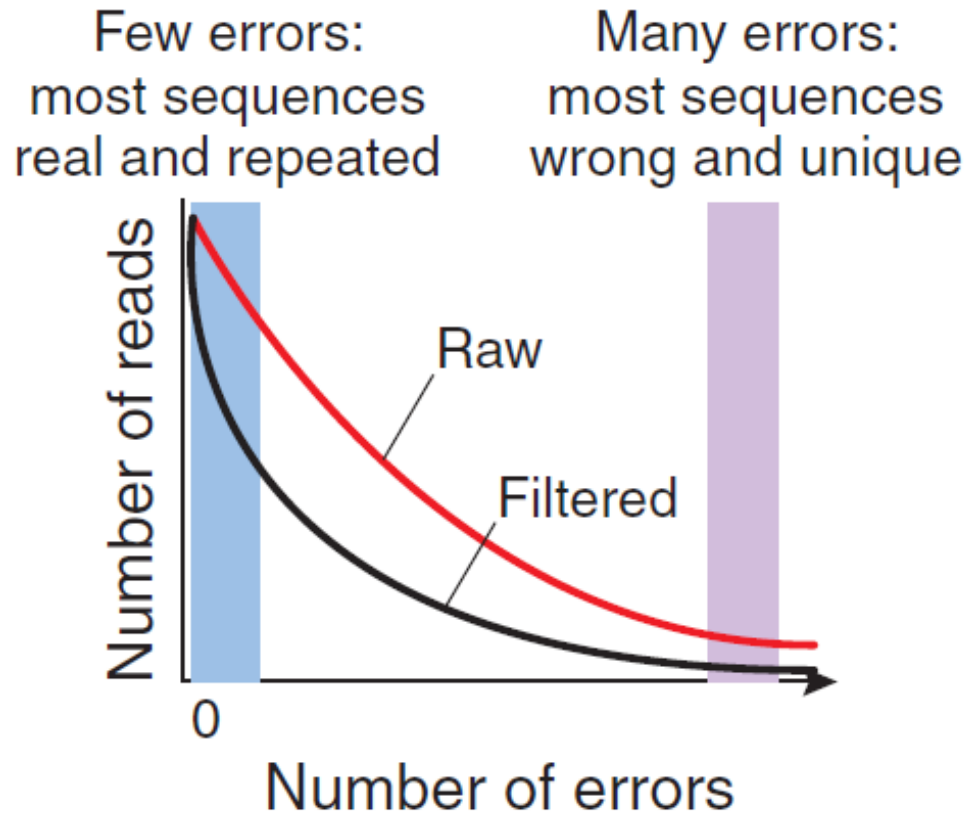


2010, Nucleic Acids Res. Claesson et al.

Filtering process



Overestimation of microbial diversity



2010, Nature Method, Reeder and Knight

Filtering effect on microbial diversity

Sample	# of reads	Subsample	Distances				
			0	gap	0.03	0.05	0.1
GM_A	10,729	1,000	586	150	196	123	51
GM_A_Flow	8,887	1,000	436		168	109	46
GM_D	4,650	1,000	567	109	227	161	85
GM_D_Flow	3,592	1,000	458		195	138	72
JM	9,929	1,000	676	50	260	195	135
JM_Flow	7,572	1,000	626		256	191	133
PM1	8,455	1,000	635	44	265	211	155
PM1_Flow	6,571	1,000	591		264	214	156
PM2	3,763	1,000	731	1	432	369	256
PM2_Flow	2,813	1,000	732		432	371	259
PM3	1,671	1,000	850	1	576	480	321
PM3_Flow	1,293	1,000	849		582	489	322

Subtle differences

Chimera slayer

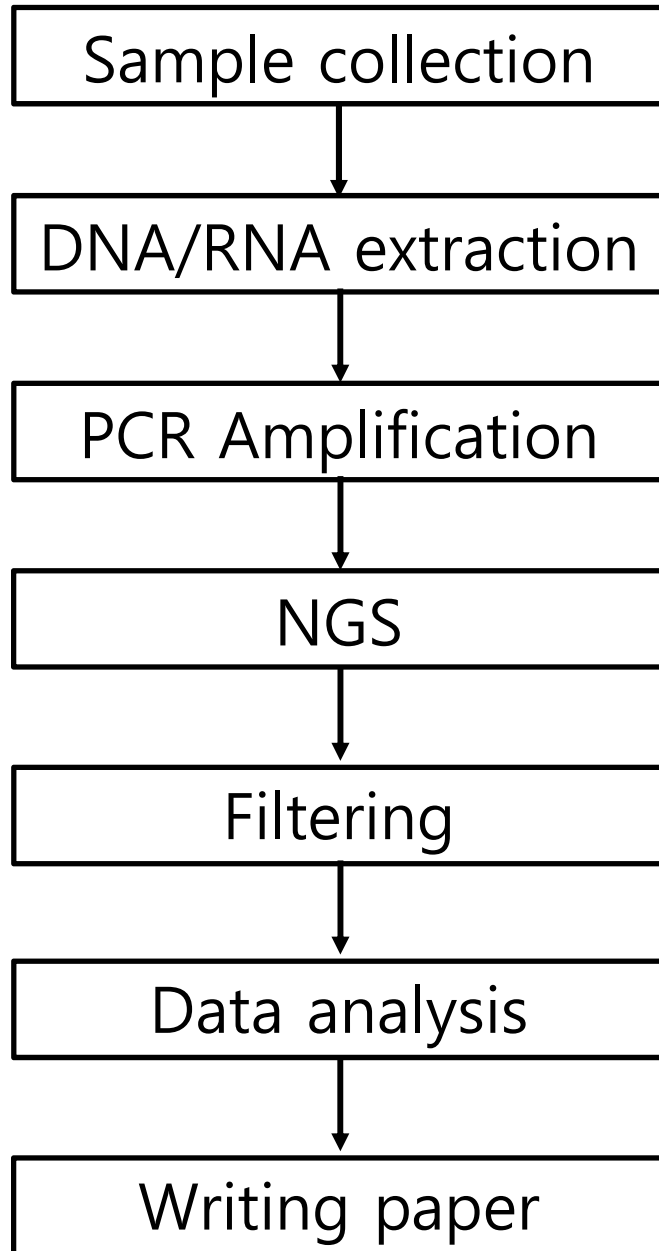
The logo for UCHIME, featuring the word "UCHIME" in a bold, orange, 3D sans-serif font. Below the letters is a faint, light-colored reflection of the text.

2011, Genome Res., Haas et al.; 2011, Bioinformatics., Edgar et al.



Step 2

Microbial structure & diversity



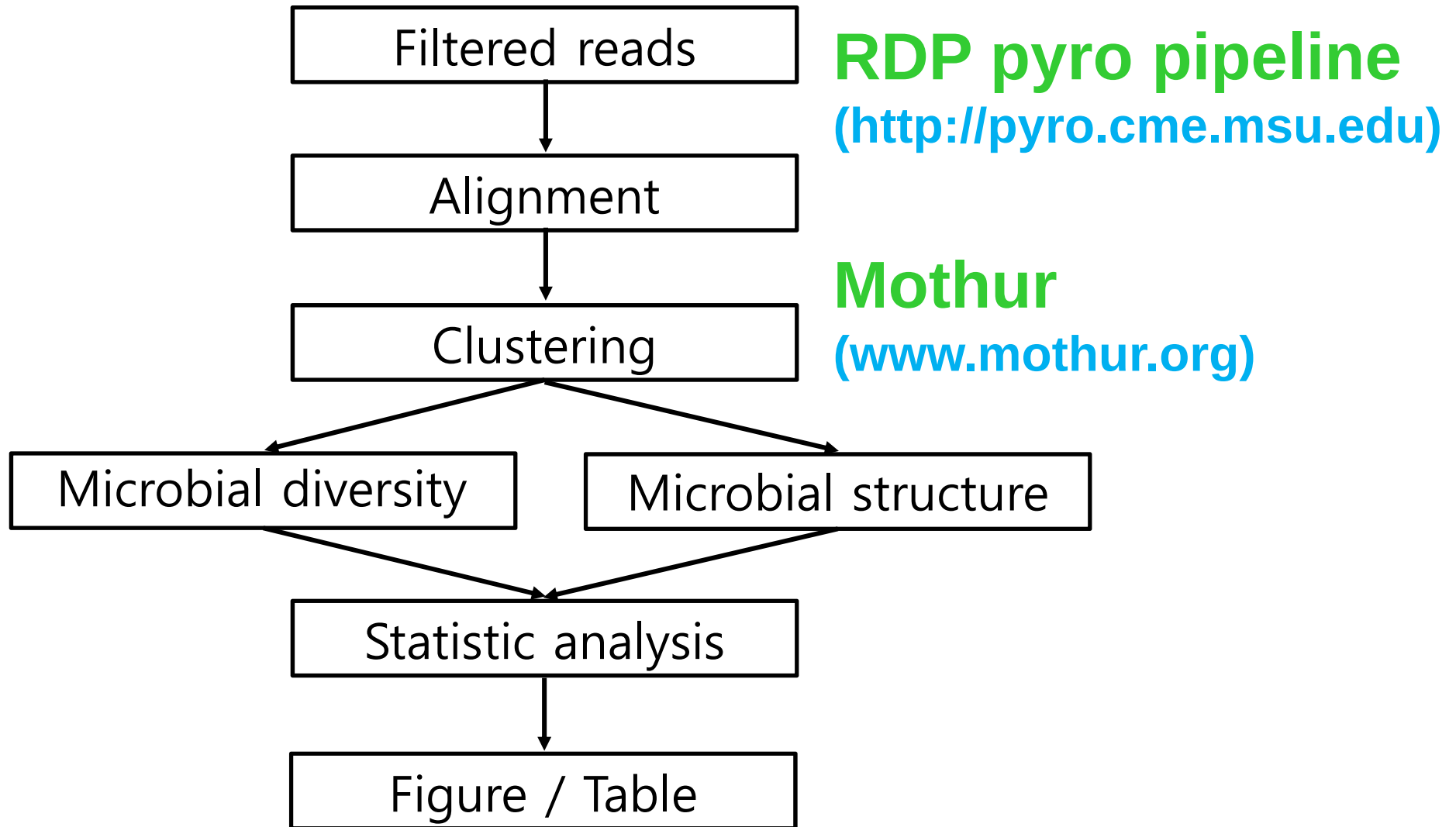
Microbial structure & Diversity

Pipeline

Statistics

Display

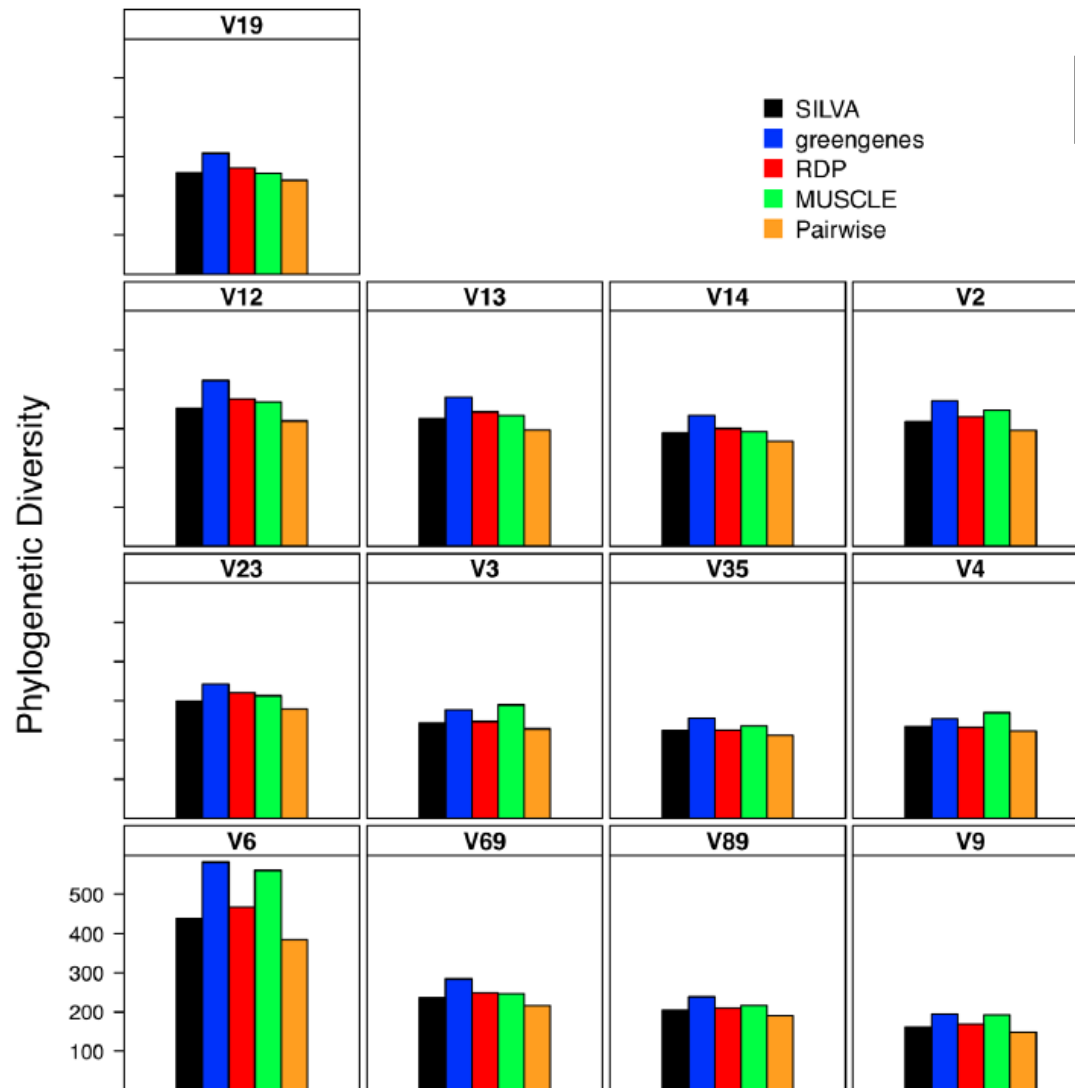
Data analysis process



Recommend

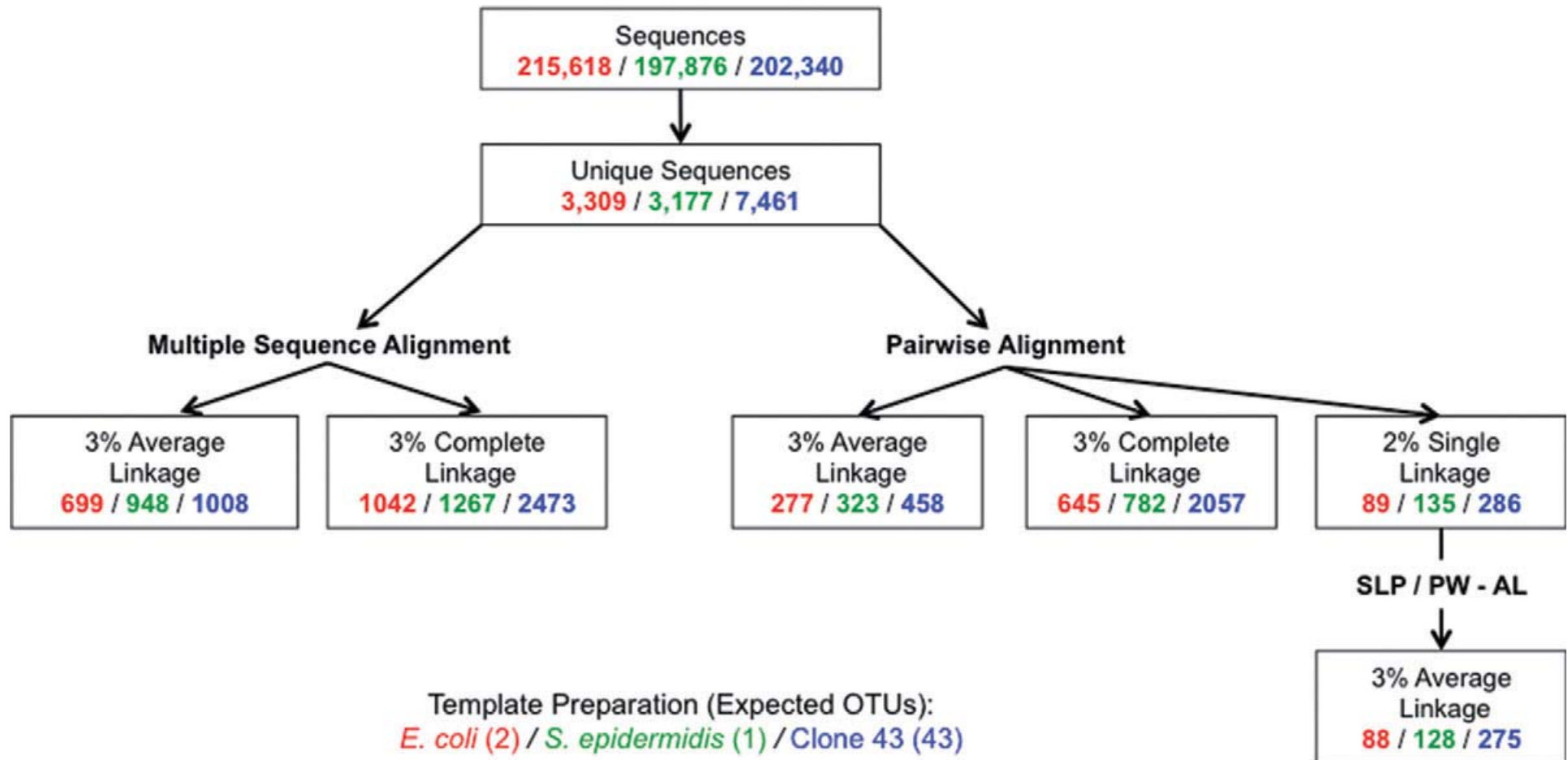
SILVA

RDP



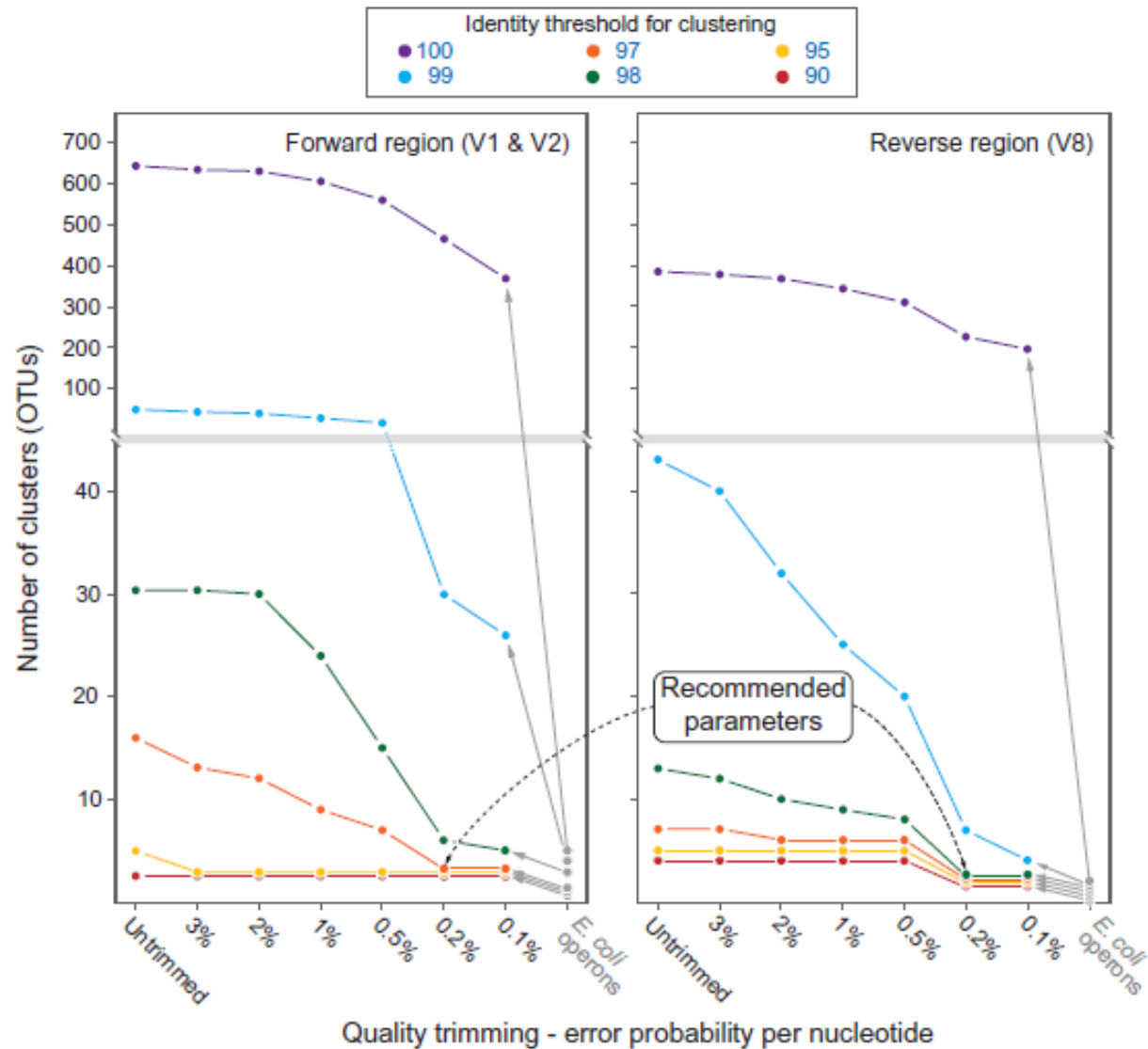
2010, PLOS Com. Bio., Schloss

Accuracy vs Time



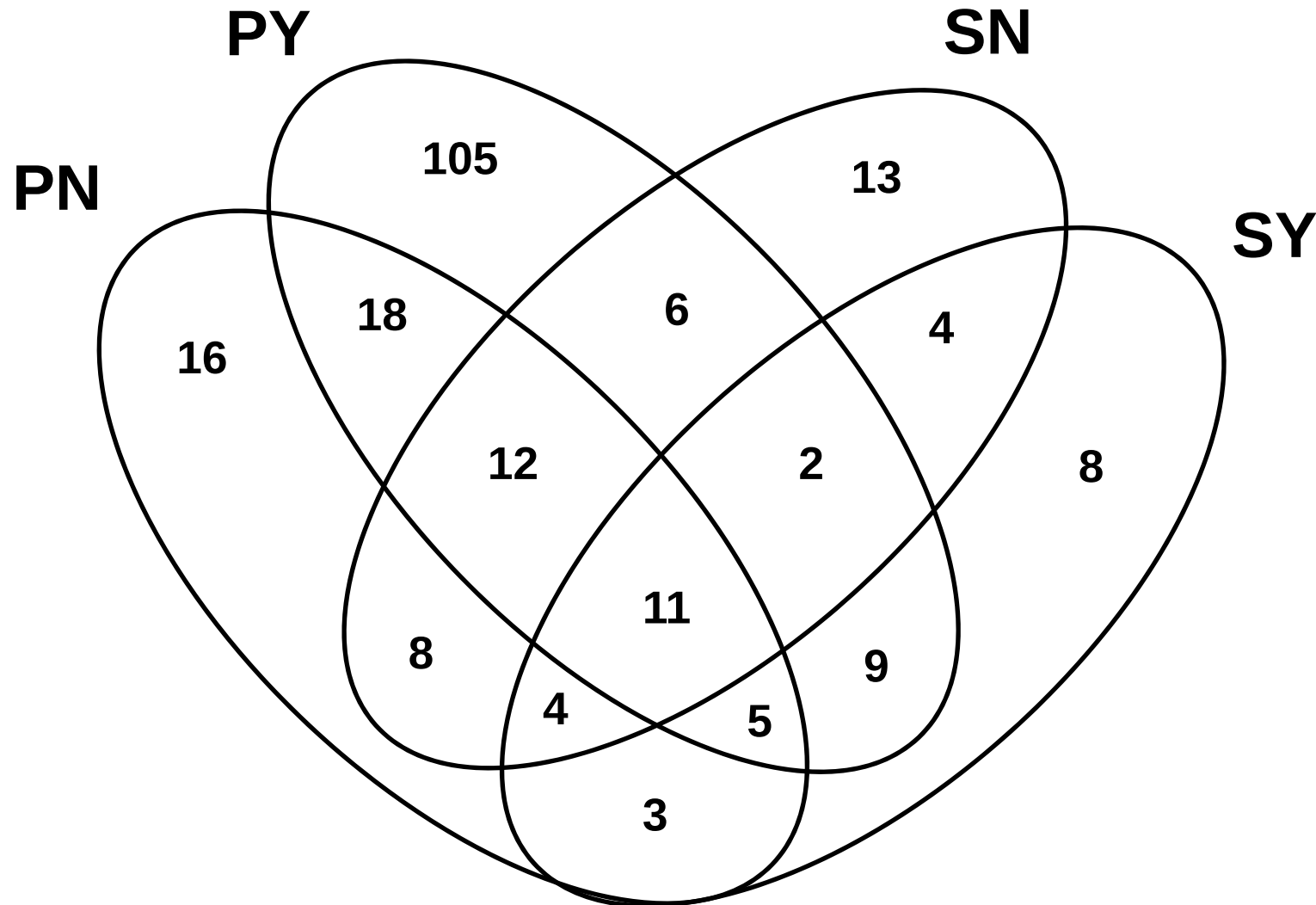
2010, *Environ. Microbiol.*, Huse et al.

3% Complete linkage clustering



2010, Environ. Microbiol., Kunin et al.

Venn Diagram



2012, J. Endo. Res., Lim et al. (submitted)

Classification

Table 3 Bacterial identification of the commonly existing anode microbes detected in this study (the identified strain is the best match with each representative population from each OTU cluster [dissimilarity cut-off<5%])

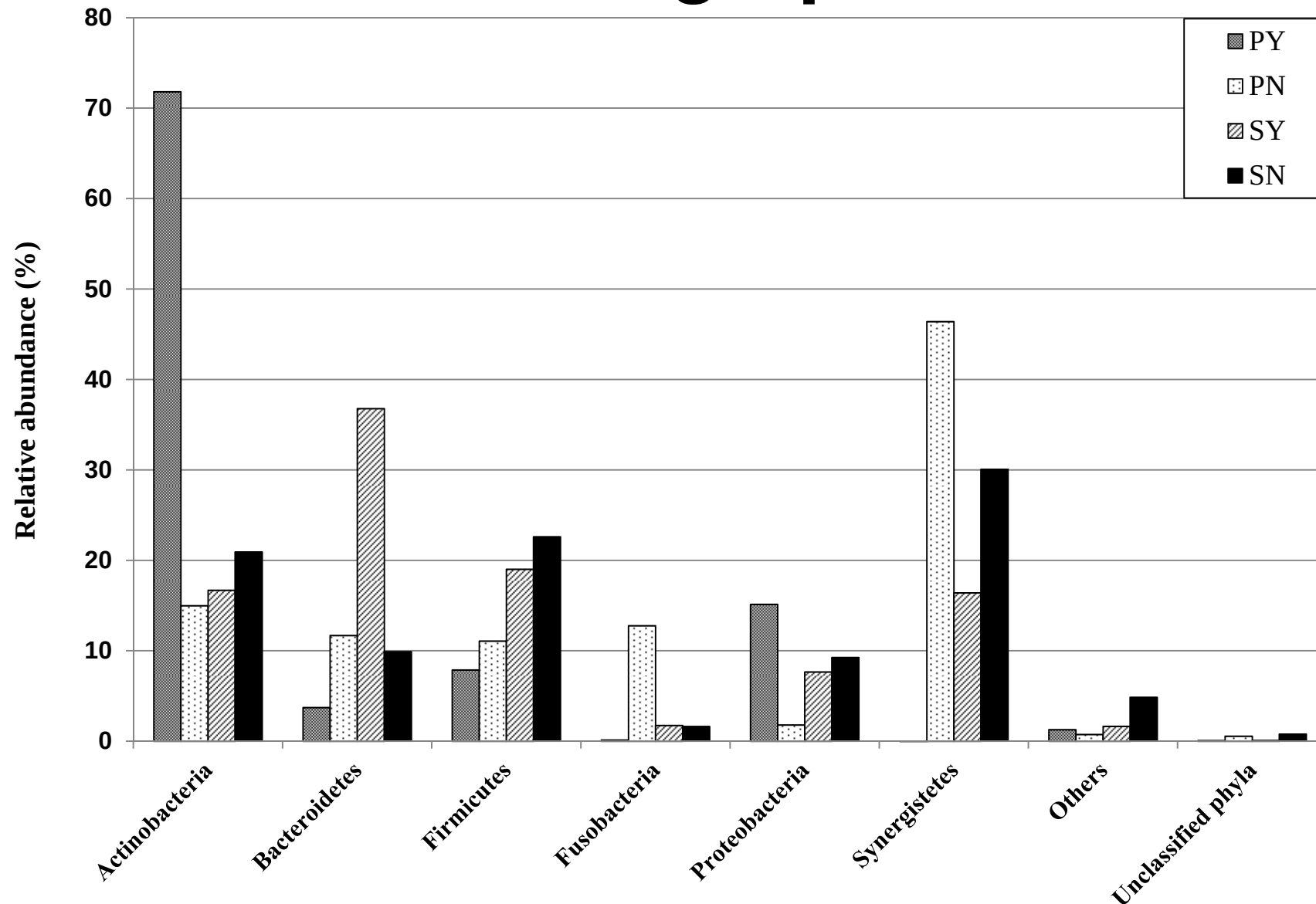
OUT ID	Accession number	RDP-II		Greengenes		NCBI BLAST	
CEAB3	GU594281	(D) <i>Geobacter psychrophilus</i> P39	81%	(D) <i>Geobacter psychrophilus</i> P35	95%	(D) <i>Geobacter psychrophilus</i> P35	95%
CEAB6	GU594282	(B) <i>Thauera</i> sp. 27	100%	(B) <i>Thauera</i> sp. 27	100%	(B) <i>Thauera</i> sp. 27	100%
CEAB8	GU594283	(B) <i>Variovorax limosa</i> EMB320	96%	(B) <i>Variovorax limosa</i> EMB320	96%	(B) <i>Polynucleobacter</i> sp. Fw70s-77	96%
CEAB9	GU594284	(G) <i>Thioflavococcus mobilis</i> 83	92%	(G) <i>Thiococcus</i> sp. AT2204	95%	(G) <i>Thiococcus pyrnigii</i> 4252	95%
CEAB11	GU594285	Unclassified bacteria	N.A.	(A) <i>Methylocystis</i> sp. KS7	81%	(A) <i>Oceanibaculum indicum</i> P24	81%
CEAB13	GU594286	(A) <i>Novosphingobium lentum</i> W-51	97%	(A) <i>Sphingomonas</i> sp. MT1	99%	(A) <i>Sphingomonas</i> sp. MT1	99%
CEAB14	GU594287	(A) <i>Aflpia</i> sp. SP17	100%	(A) <i>Aflpia</i> sp. SP17	100%	(A) <i>Aflpia</i> sp. SP17	100%
CEAB15	GU594288	(F) <i>Fuzibacter</i> sp. SA1	95%	(F) <i>Fuzibacter</i> sp. SA1	95%	(F) <i>Fuzibacter</i> sp. SA1	95%
CEAB17	GU594289	(G) <i>Pseudomonas mendocina</i> PC12	99%	(G) <i>Pseudomonas mendocina</i> PC12	99%	(G) <i>Pseudomonas mendocina</i> PC12	99%
CEAB18	GU594290	Unclassified bacteria	N.A.	(C) <i>Dehalococcoides</i> sp. BHI80-15	75%	(D) <i>Sorangium cellulosum</i> M5	85%
CEAB19	GU594291	(G) <i>Acinetobacter</i> sp. DZ0503SBS4	98%	(G) <i>Acinetobacter</i> sp. DZ0503SBS4	98%	(G) <i>Acinetobacter</i> sp. DZ0503SBS4	98%
CEAB21	GU594292	(F) <i>Clostridium lituseburense</i>	93%	(F) <i>Clostridium</i> sp. C01-2409	97%	(F) <i>Clostridium lituseburense</i>	96%

The letter in parenthesis indicates the identified phylum for each OUT group. The percent number indicates sequence similarity between the best-matched database sequence and the representative sequence of each OUT group

Type strain identification : Ez-Taxon

2010, Appl. Microbial. Biotech., Lee et al.

Bar graph



2012, J. Endo. Res., Lim et al. (submitted)

Statistics analysis



Open
source
statistical
computing.

+

VEGAN

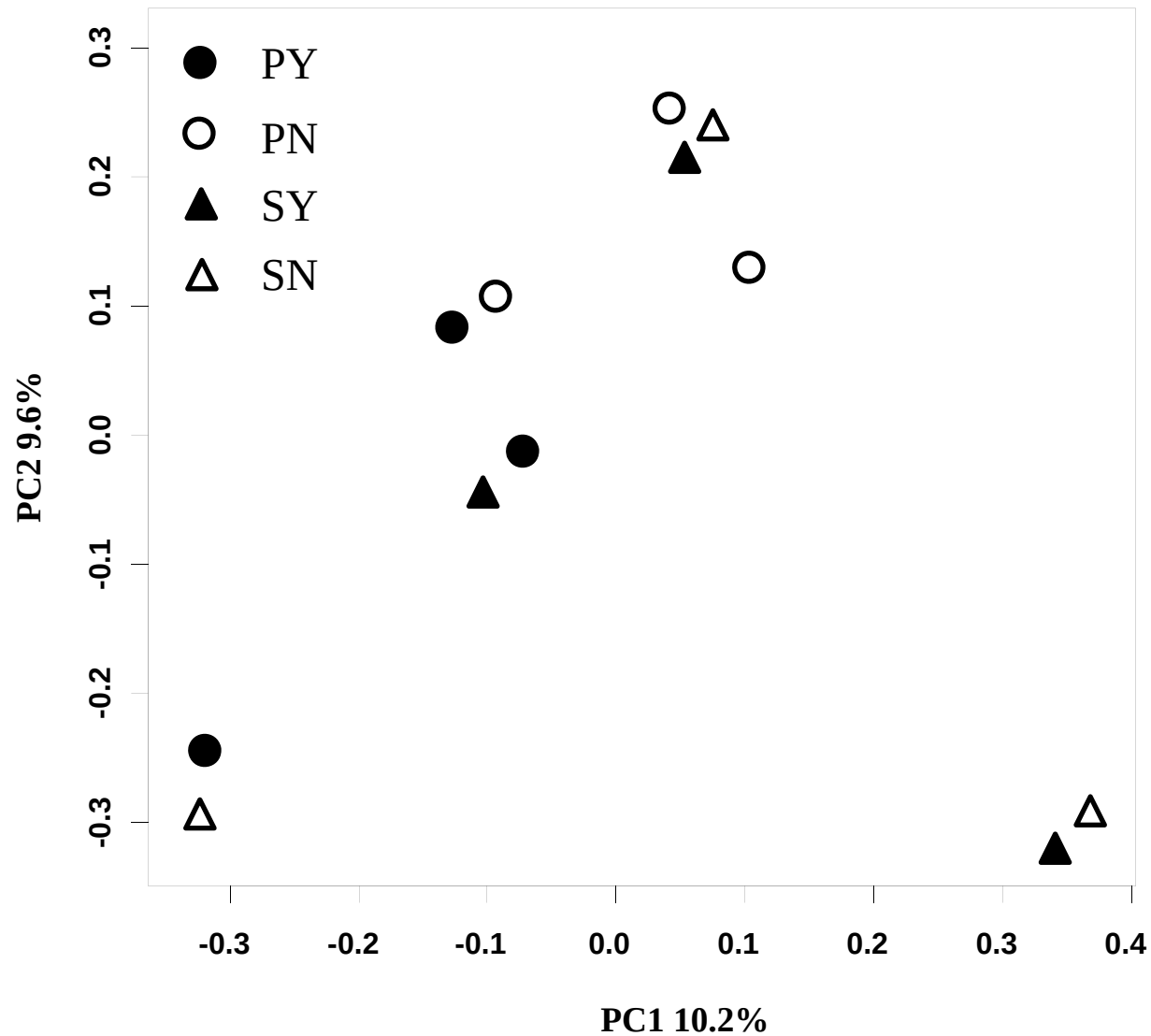
Pirate for
Statistics!

BiodiversityR

Contribute at cran.r-project.org

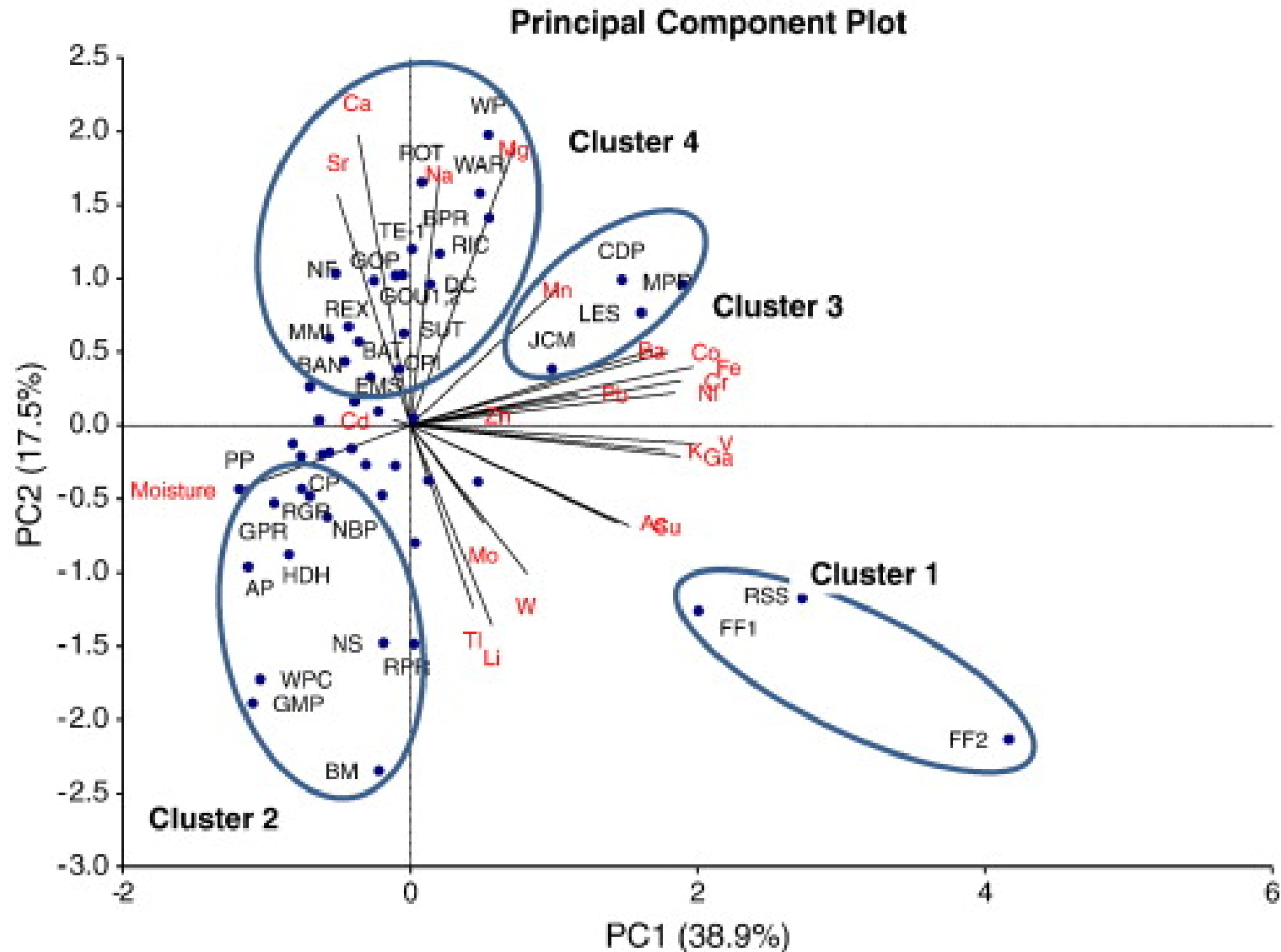
<http://www.r-project.org>

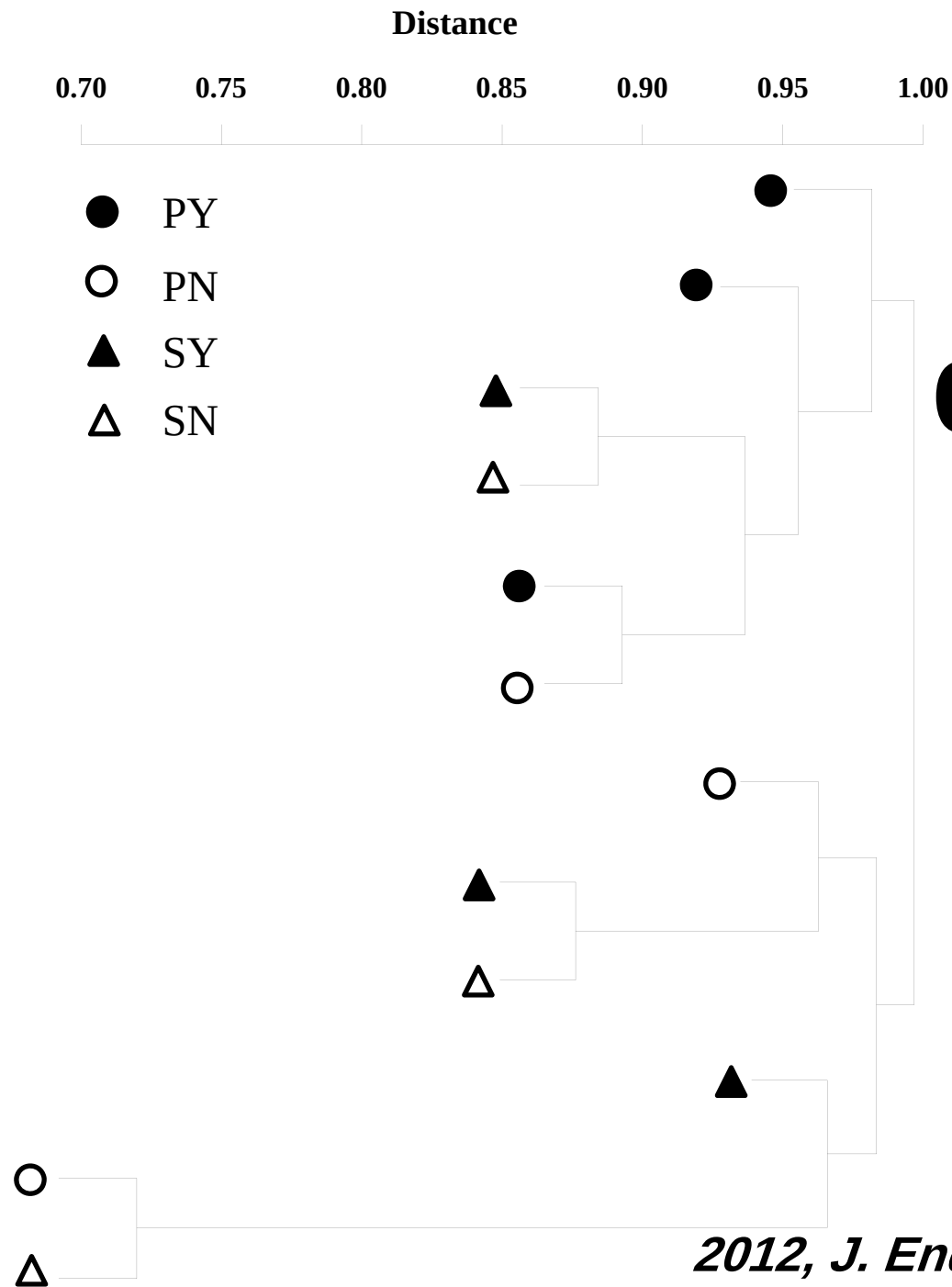
PCA analysis



2012, J. Endo. Res., Lim et al. (submitted)

PCA analysis with metadata

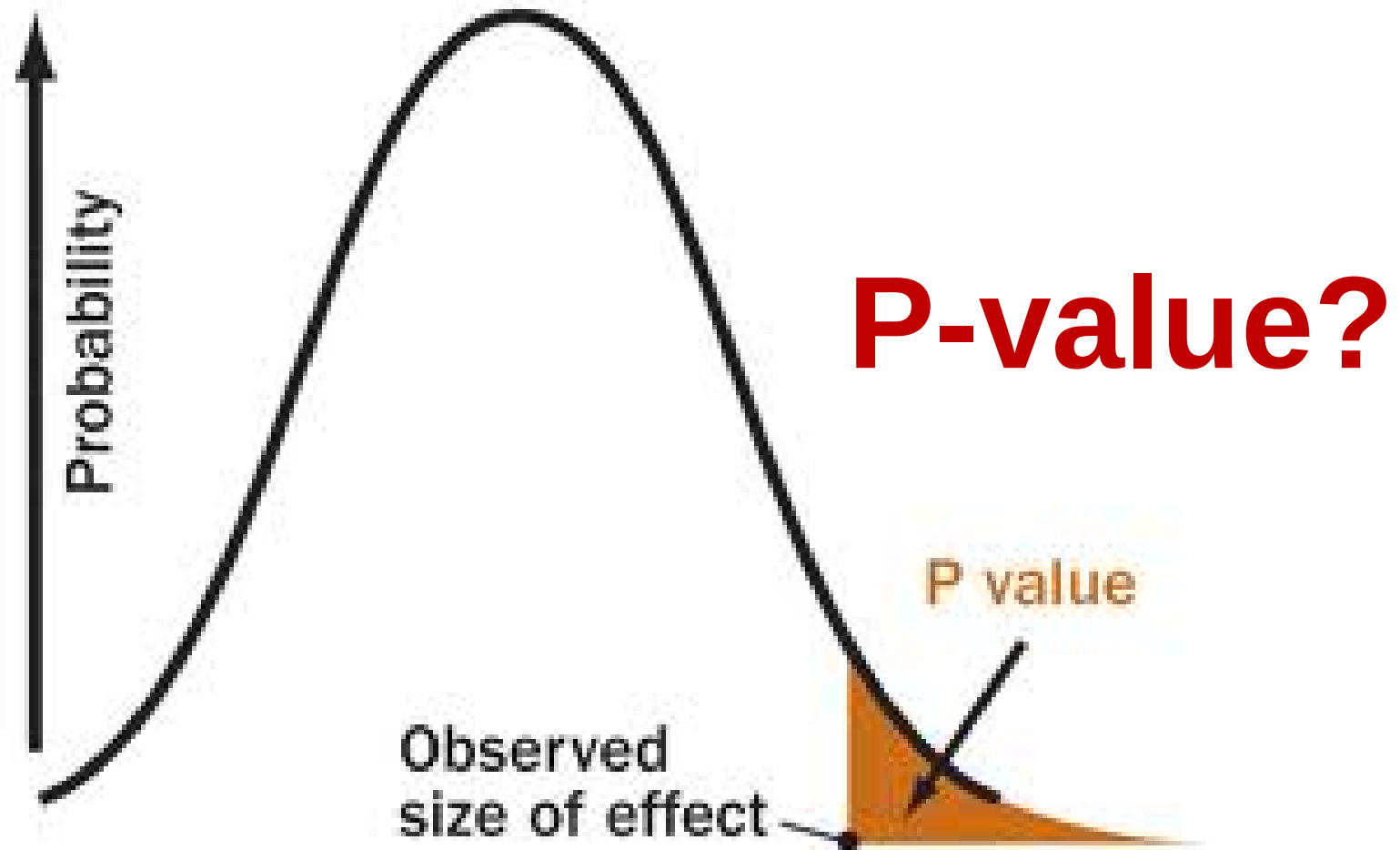




UMPGA dendrogram

2012, J. Endo. Res., Lim et al. (submitted)

Statistics analysis

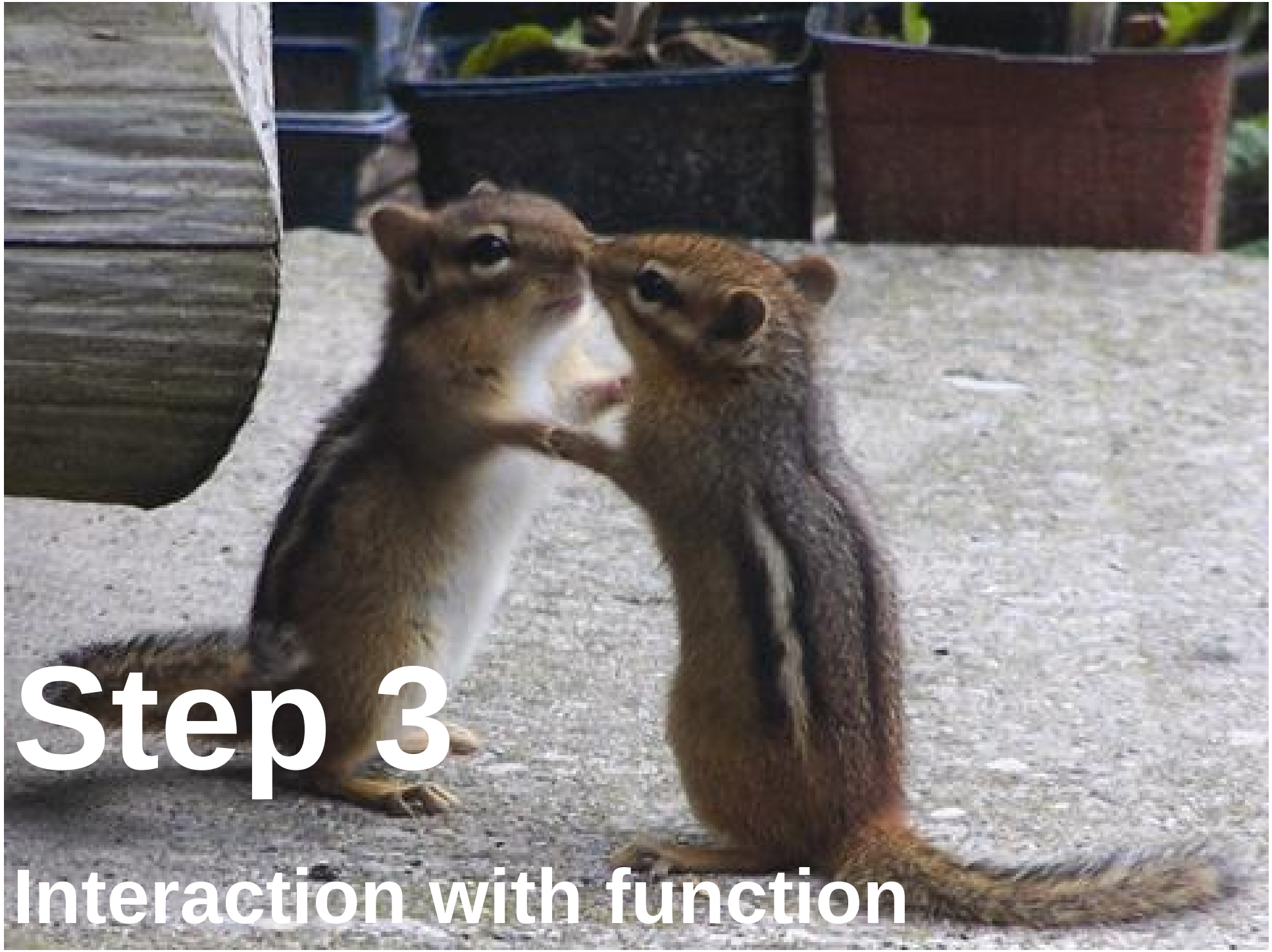


Basic significance

R & library compare (RDP)

Parsimony test (P-test)

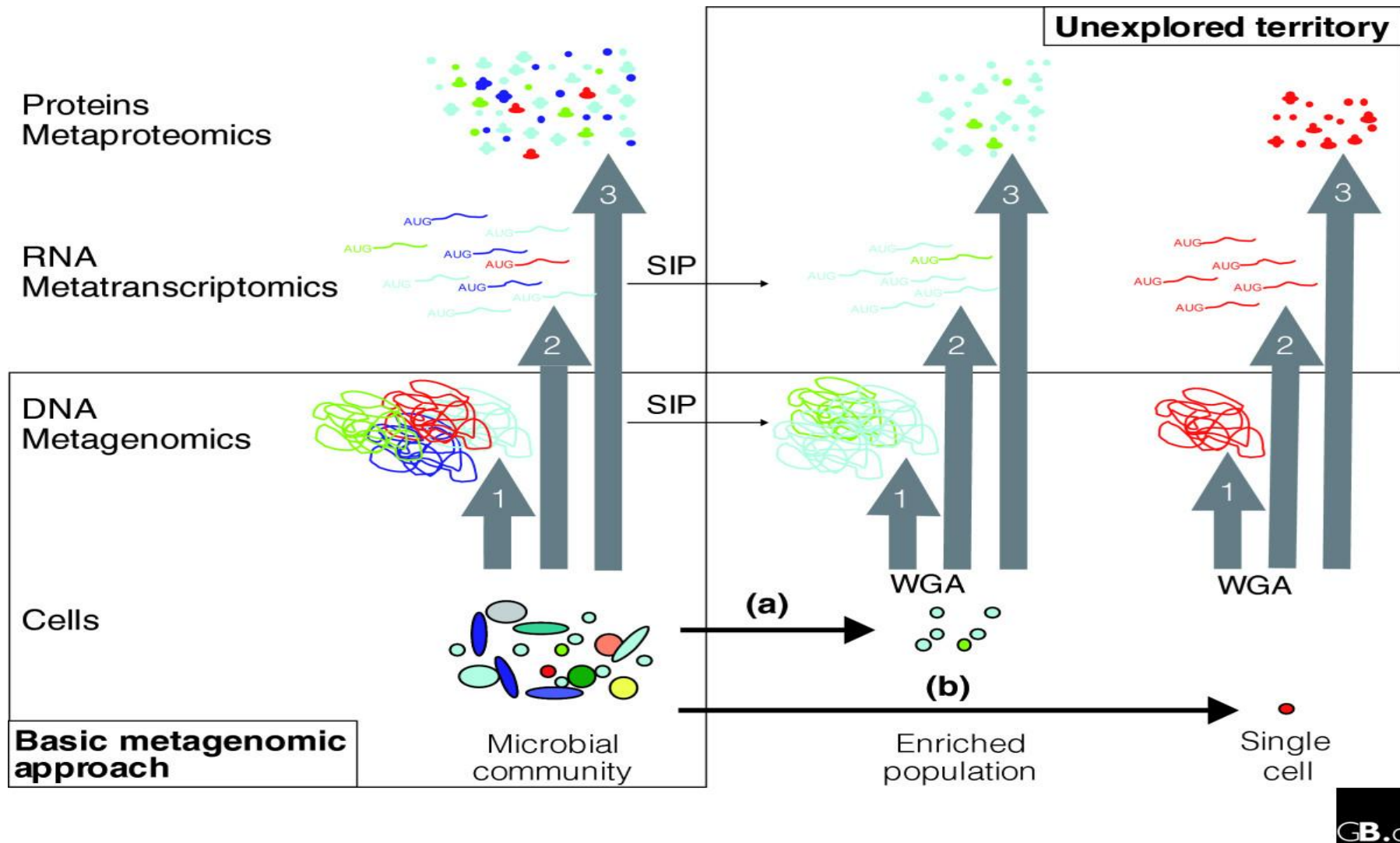
Libshuff, ANOVA and HOMOVA

A photograph of two chipmunks on a concrete surface. The chipmunk on the left is standing on its hind legs, facing the other chipmunk. The chipmunk on the right is also standing on its hind legs, reaching up with its front paws towards the face of the first chipmunk. They appear to be in a social interaction. In the background, there are some potted plants and a wooden structure.

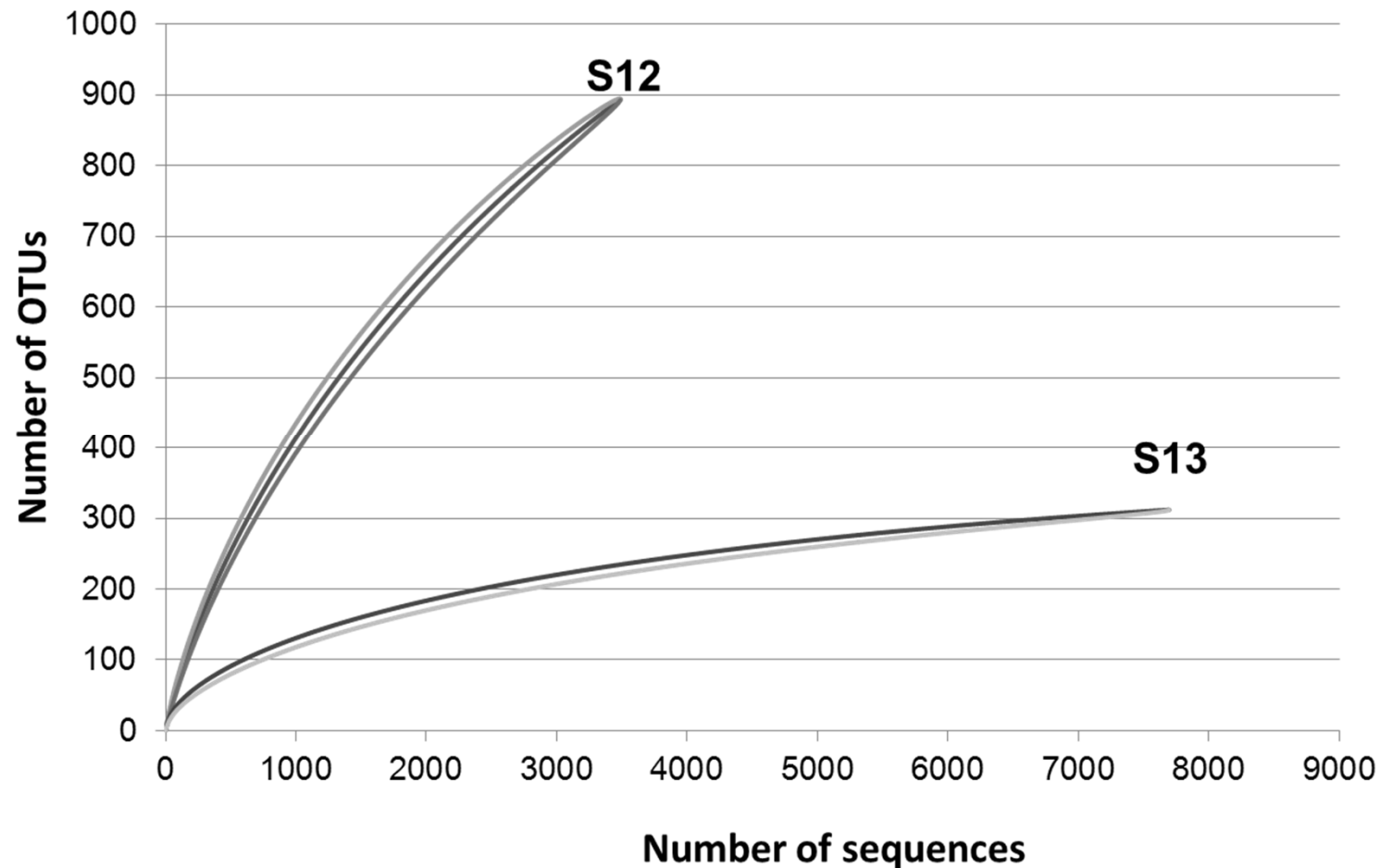
Step 3

Interaction with function

Function with microbial structure

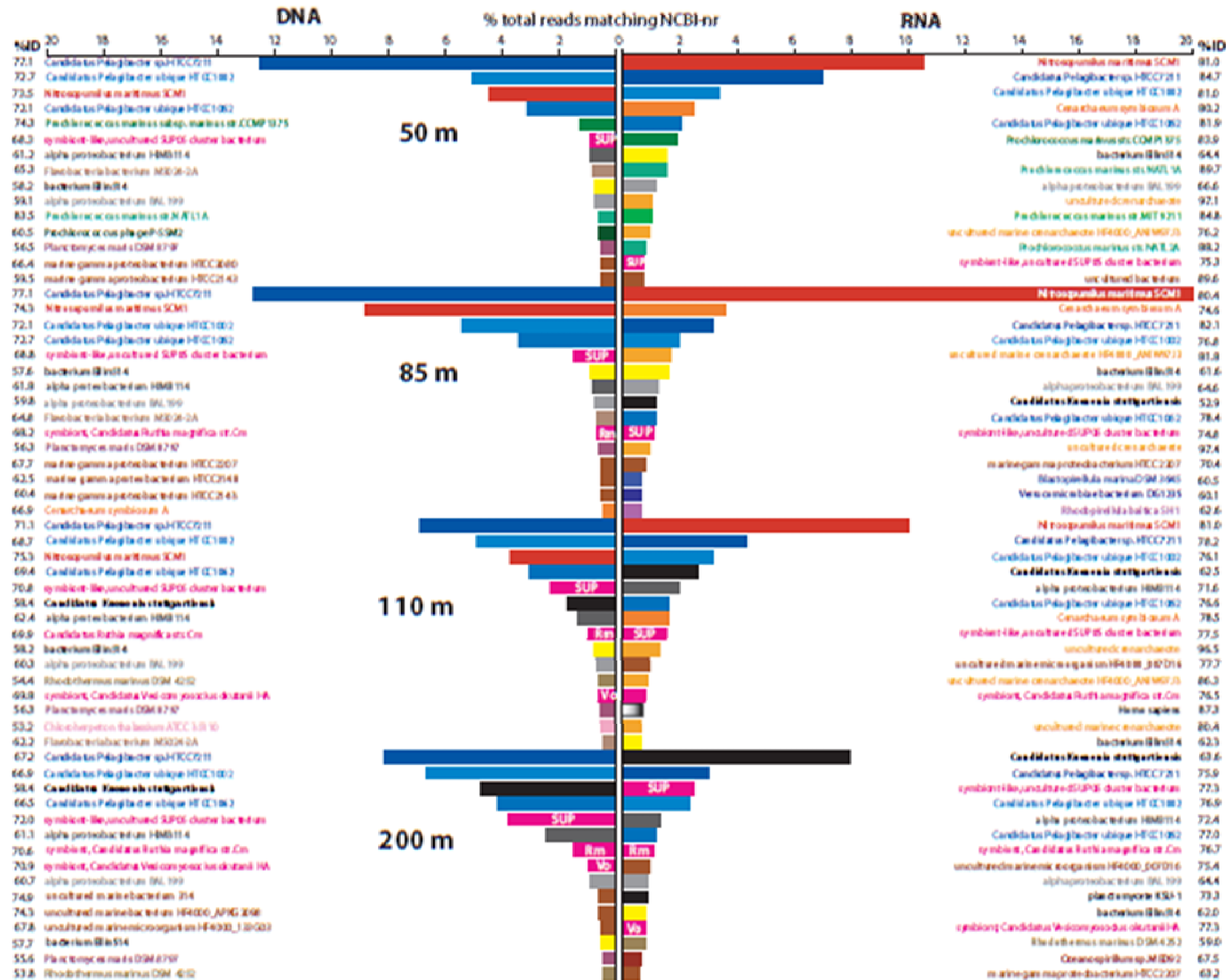


Stable isotope probing (SIP)



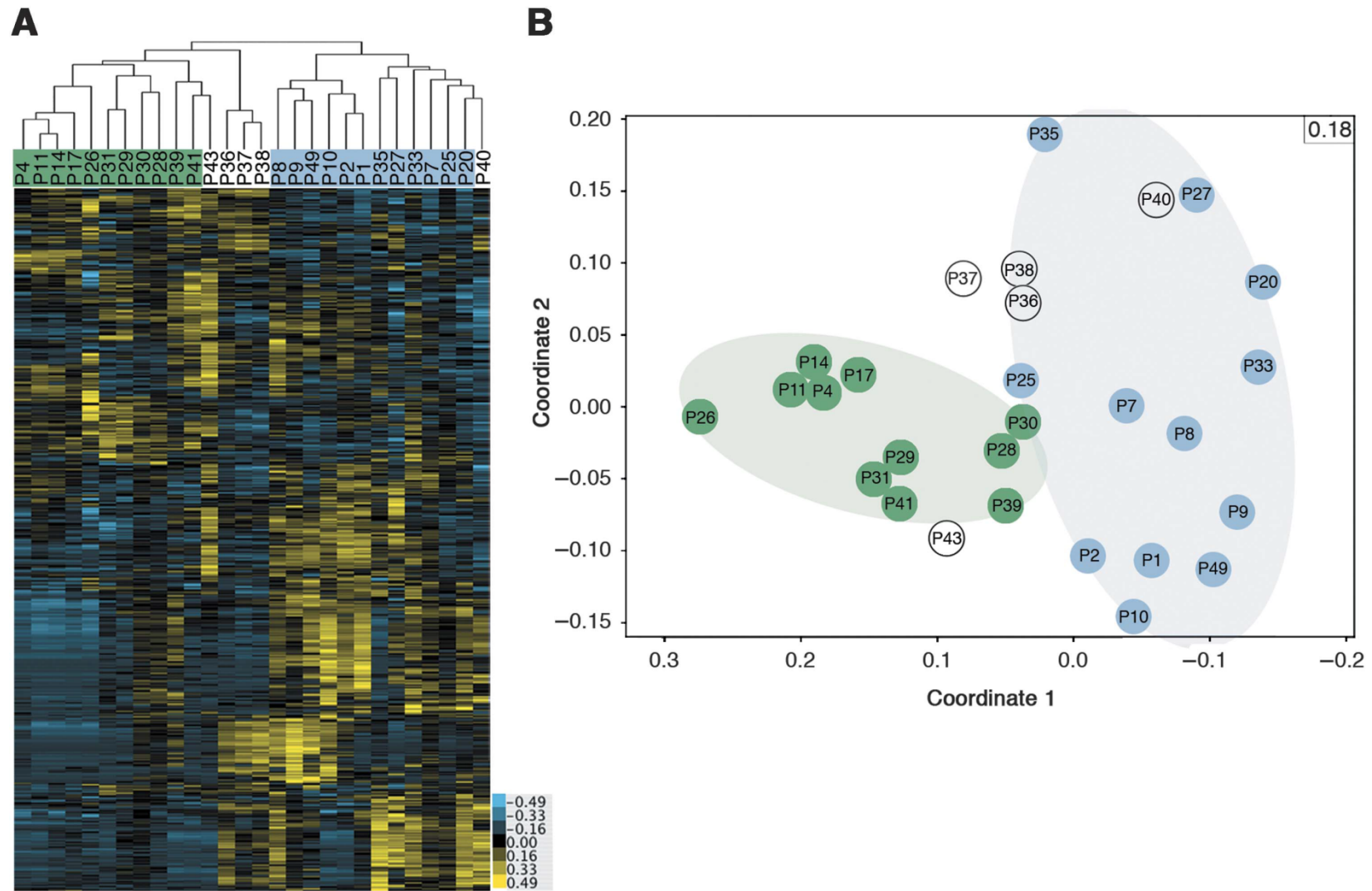
2011, Appl. Environ. Microbiol., Lee et al.

Metagenome & Metatranscriptome



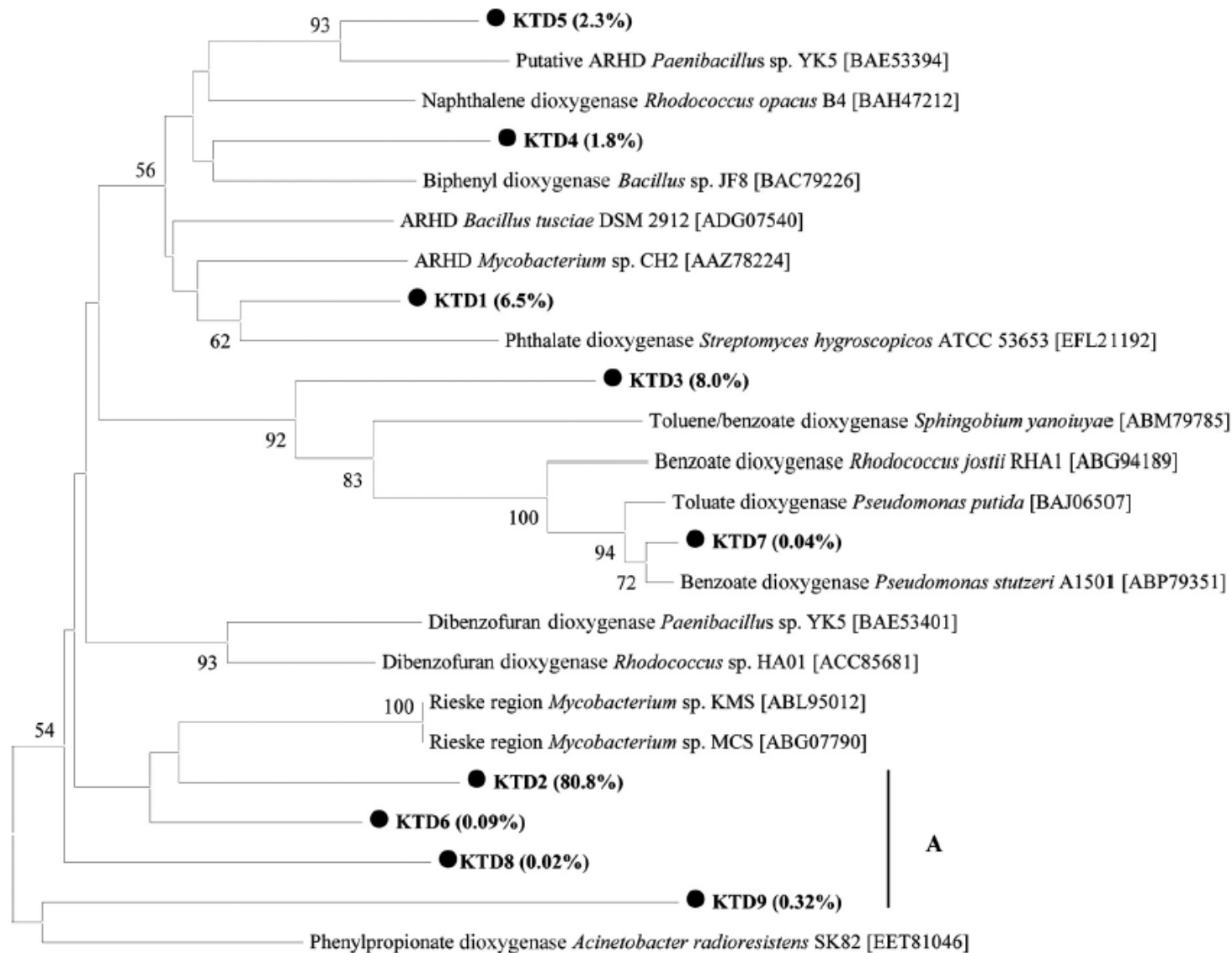
2011, Environ. Microbiol., Stewart et al.

Metaproteomics



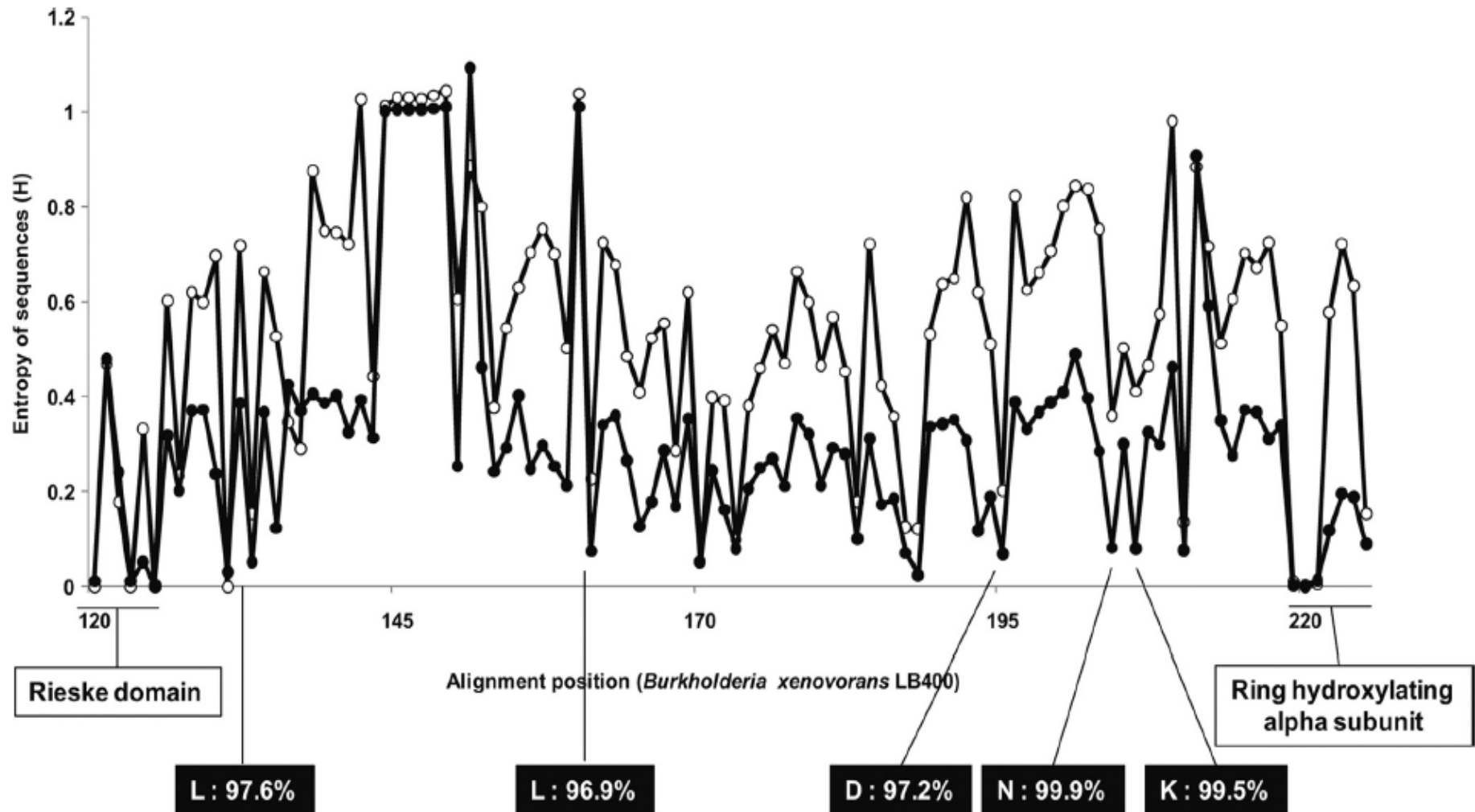
2010, Mol. Sys. Biol., Mueller et al.

Functional gene analysis 1



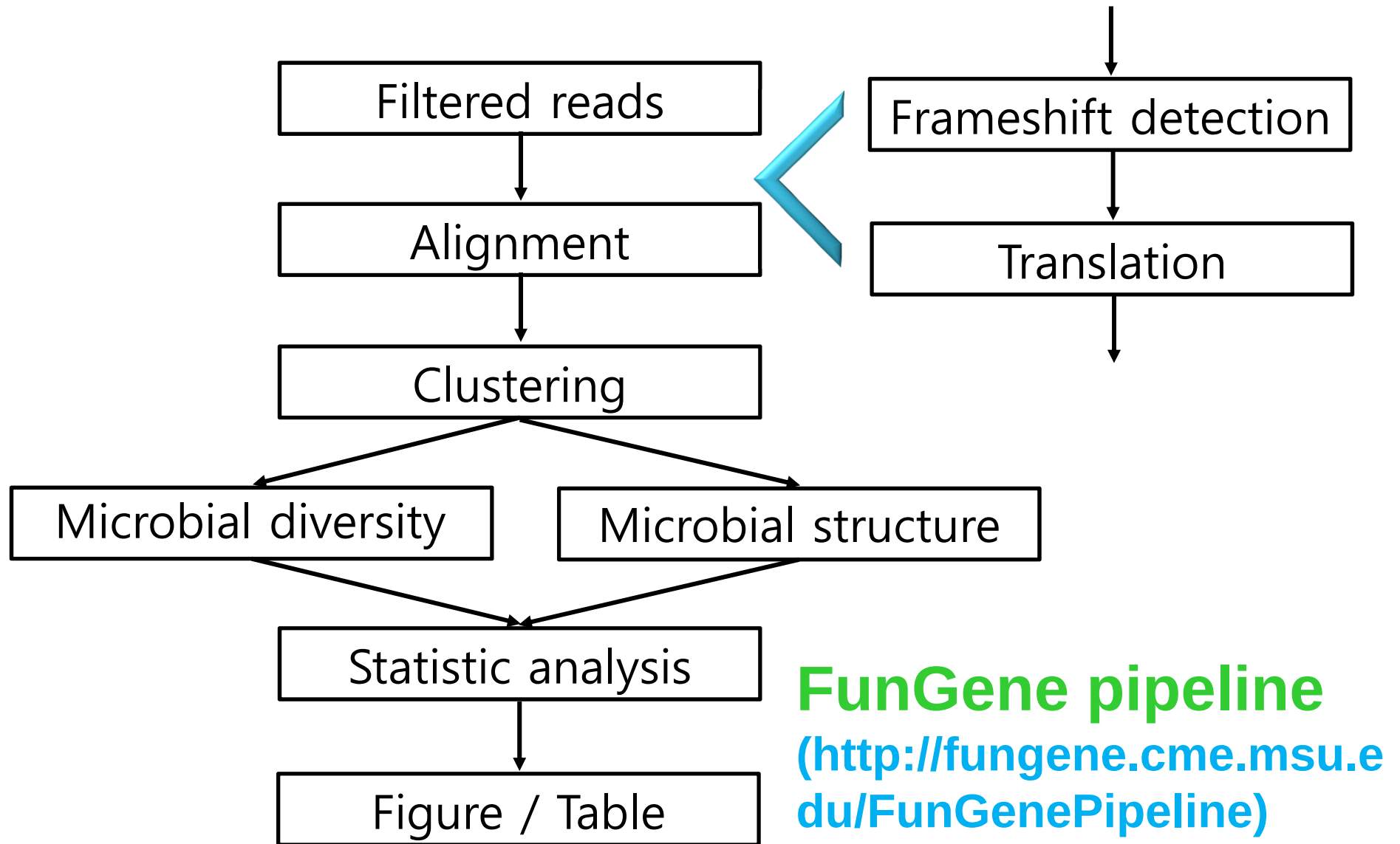
2011, Appl. Environ. Microbiol., Lee et al.

Functional gene analysis 1



2011, Appl. Environ. Microbiol., Lee et al.

Data analysis process



Antibiotic resistances (6)

Biodegradation (18)

Biogeochemical cycles (26)

Phylogenetic markers (11)

Plant pathogenicity (2)

11 genes

nifH, nirK

- 1. Can we determine the true microbial diversity (phylogenetically and genetically)?**
- 2. Is there a core microbiome?**
- 3. Does structure impact ecological function? If so, how?**



3

Steps of

Microbial analysis

1. **Sample preparation & filtering**
2. **Microbial structure & diversity**
3. **Interaction with function**

Acknowledgement

WCU Center for Green Metagenomics @ Yonsei University

- Prof. Joonhong Park
- Jaejin Lee
- Jangho Lee

Center for Microbial Ecology @ Michigan State University

- Prof. James M. Tiedje

Ribosomal Database Project @ Michigan State University

- Prof. James Cole
- Qiong Wang

Marine Biological Laboratory @ Woods Hole

- Woo Jun Sul



The background of the slide is a dark blue field filled with a complex, glowing pattern of white and light blue lines. These lines form a dense, interconnected web that resembles a digital network or a stylized DNA double helix. The pattern is more concentrated on the right side, where it appears as a bright, swirling vortex, and fades into a darker, more grid-like structure on the left.

Thank you.

Contact : blurple@live.co.kr